



Optical Microscopy

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Outline

- Conventional Light Microscopy

Brightfield, phase & interference contrast

- Fluorescence Microscopy
- Confocal and Two-Photon Microscopy
- TIRF Microscopy
- Advances In Fluorescence Microscopy

Microscopy Resources

- Web:

www.microscopyU.com (Nikon)

Micro.magnet.fsu.edu

www.olympusmicro.com (Olympus)

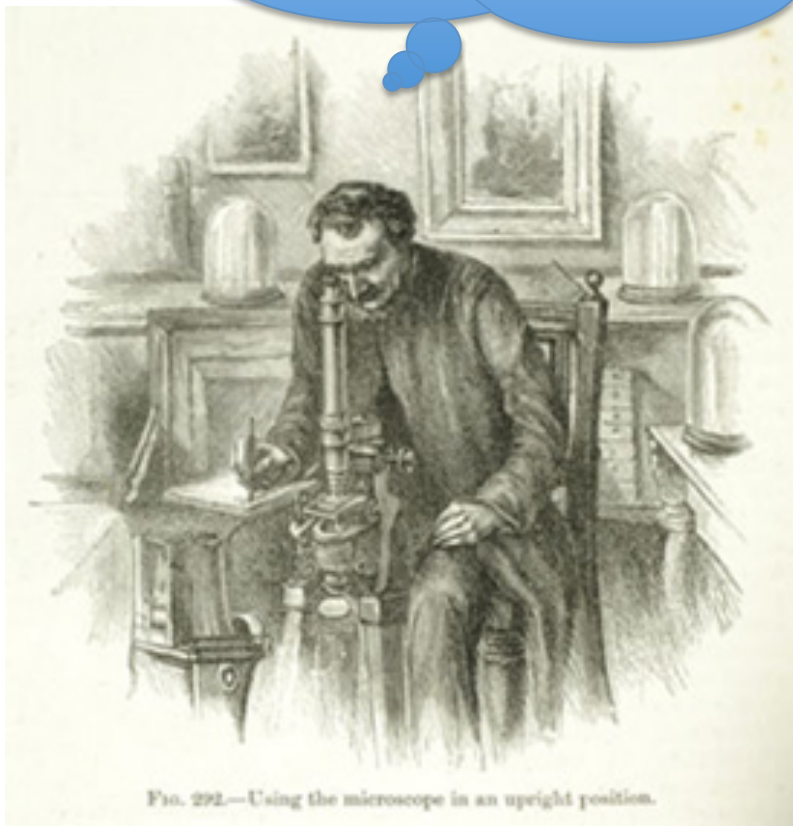
- Papers (will be uploaded to course website)

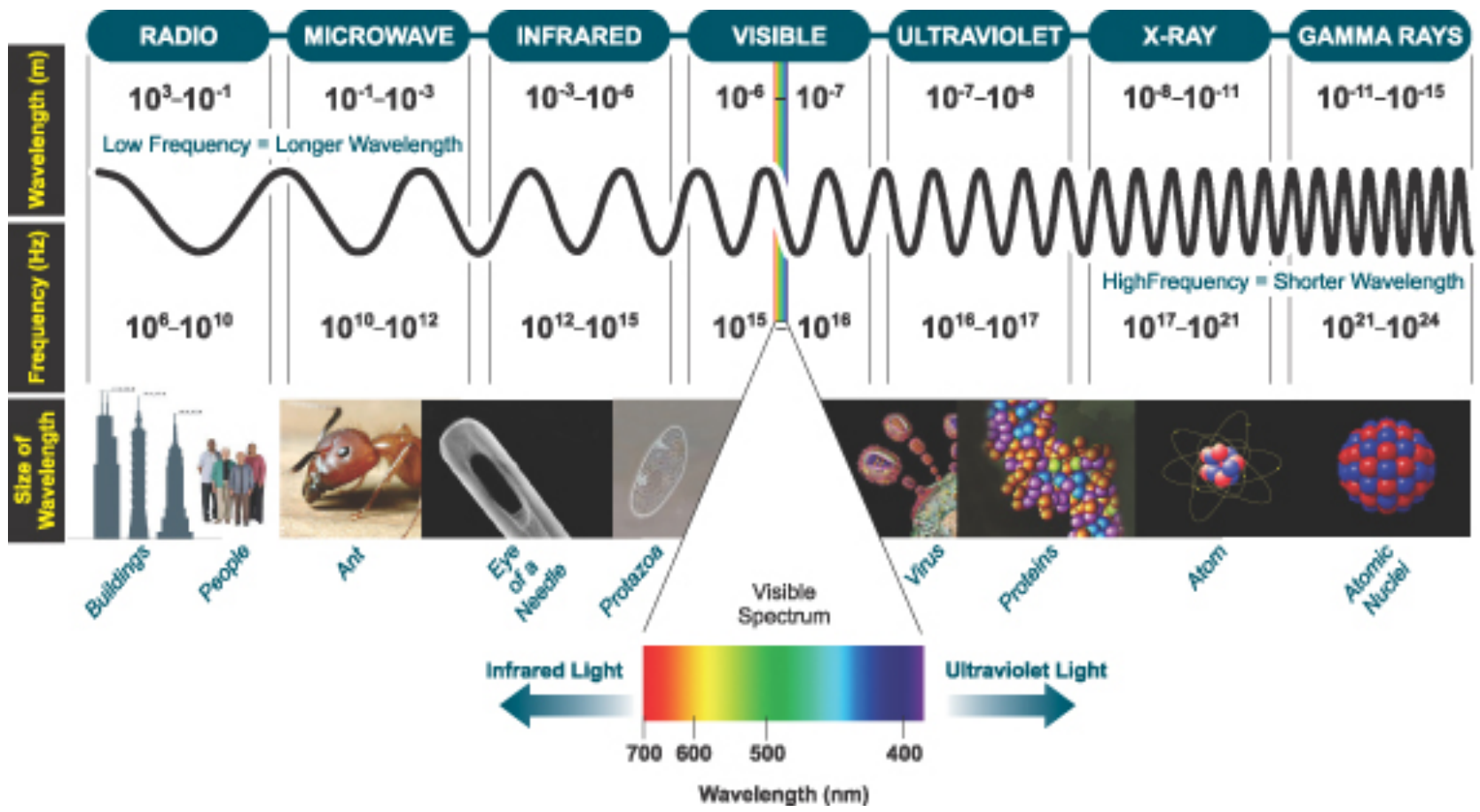
Lecture slides are based on the above resources...

Additional resources (Molecular probes/Invitrogen)

Some notes and slides adapted/modified from Prof. Aryeh Weiss
(BIU@Israel)

What is the Physical Nature of Light?
What is Fluorescence?

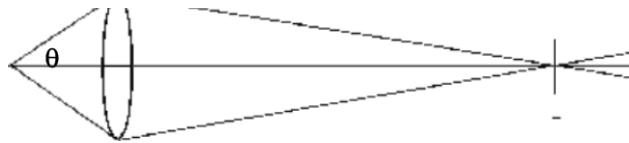




Andor Technology

Descriptions for the Properties of Light

Most Basic: Ray diagrams.

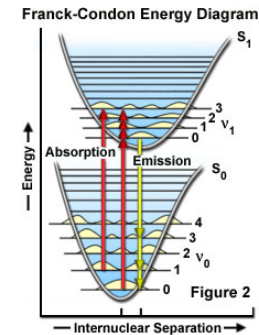


Classical: Electromagnetic waves.

$$\frac{\partial^2 \mathbf{E}}{\partial t^2} - c_0^2 \cdot \nabla^2 \mathbf{E} = 0$$

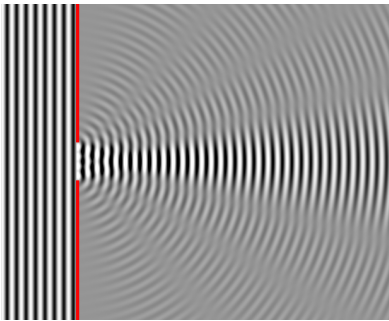
$$\frac{\partial^2 \mathbf{B}}{\partial t^2} - c_0^2 \cdot \nabla^2 \mathbf{B} = 0$$

Light-Matter Interaction: Quantum Entities – Photons.



The Duality of Light: **What is the Physical Nature of Light?**

Light behaves like... Waves
(Diffraction, Interference, refraction)

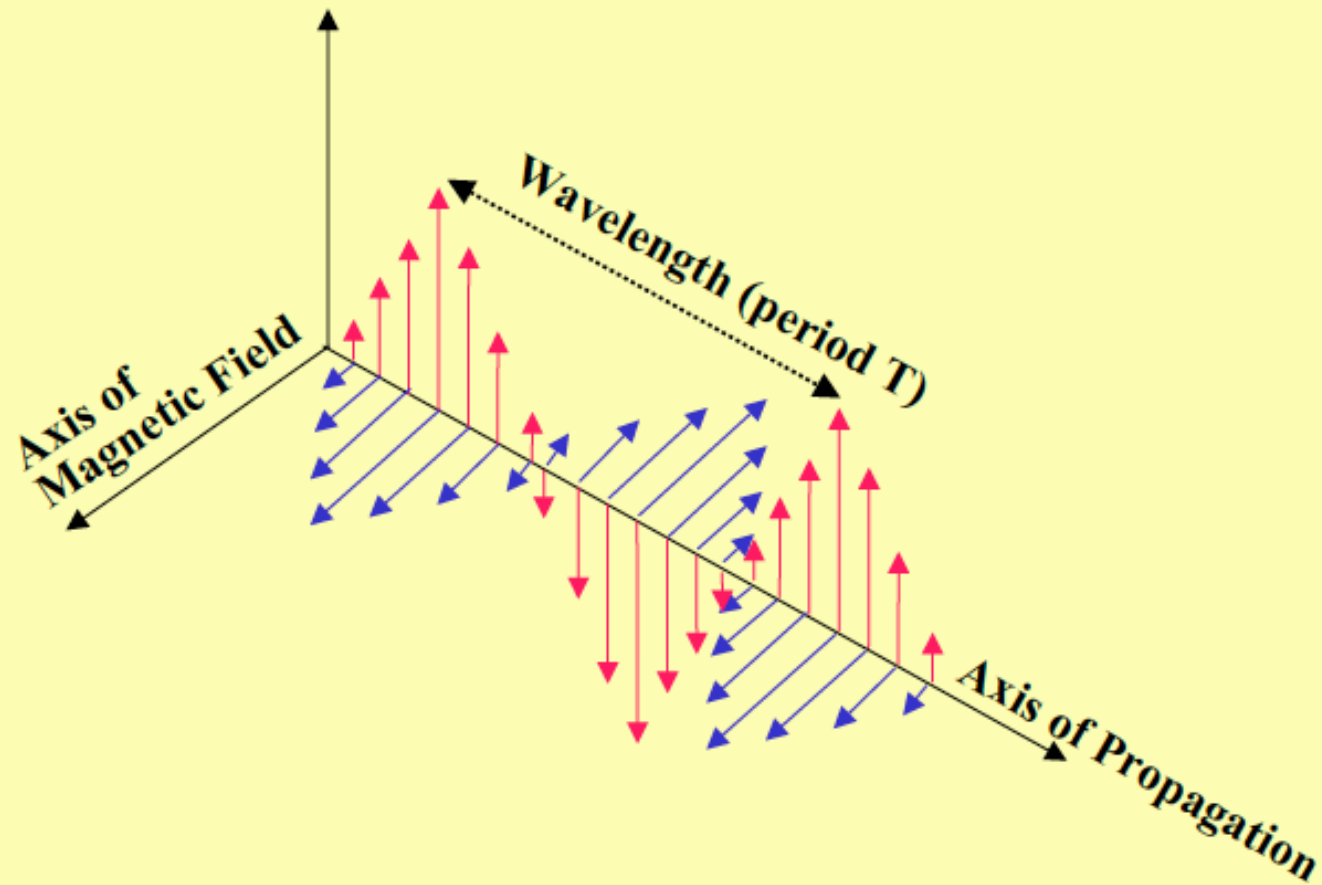


Newton, Huygens, Fresnel, Young, Maxwell

Maxwell's Equations
(Electromagnetic Waves)

$$\frac{\partial^2 \mathbf{E}}{\partial t^2} - c_0^2 \cdot \nabla^2 \mathbf{E} = 0$$
$$\frac{\partial^2 \mathbf{B}}{\partial t^2} - c_0^2 \cdot \nabla^2 \mathbf{B} = 0$$

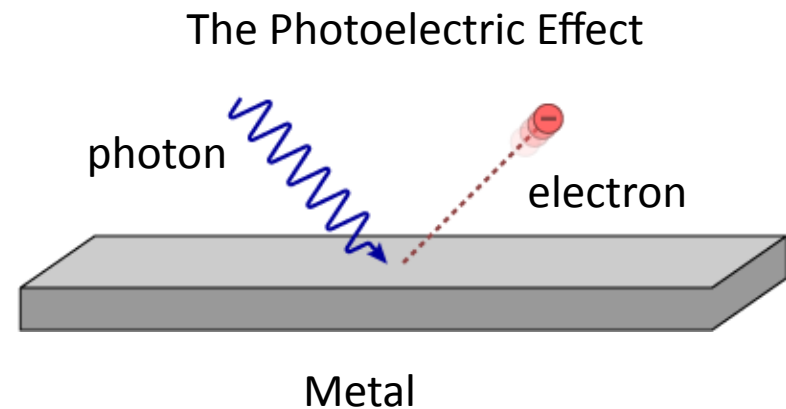
ELECTROMAGNETIC WAVE



The Duality of Light: **What is the Physical Nature of Light?**

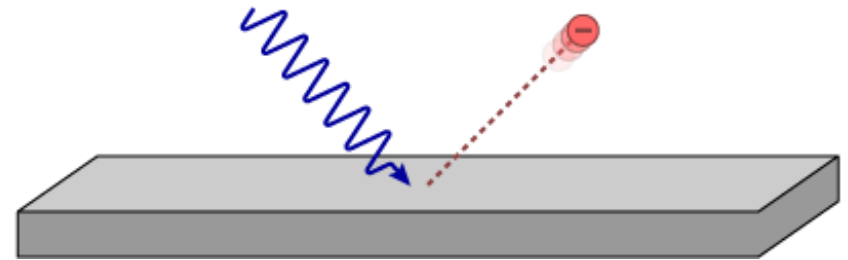
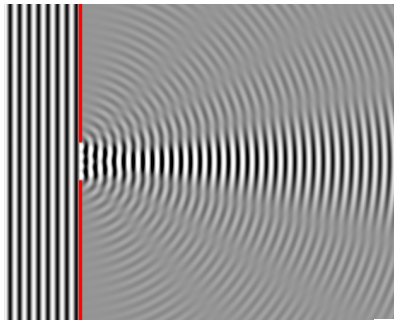
Light behaves like.... Particles
(Discrete Quantities of Energy- light quantas or photons)

Hertz, Thomson, Lenard, Einstein



The Duality of Light

Light has a duality, wave **AND** particle



"Once and for all I want to know what I'm paying for. When the electric company tells me whether light is a wave or a particle I'll write my check."

The Duality of Light

Equations that describe the properties of light:

Wave equations: (frequency, amplitude).

$$\Psi(x,t) = A * \exp(i(\kappa x + \omega t))$$

$$\kappa = 2\pi/\lambda$$

$$\omega = 2\pi\nu$$

Energy of Photons:

$$E = h\nu = hc/\lambda$$

ν -frequency

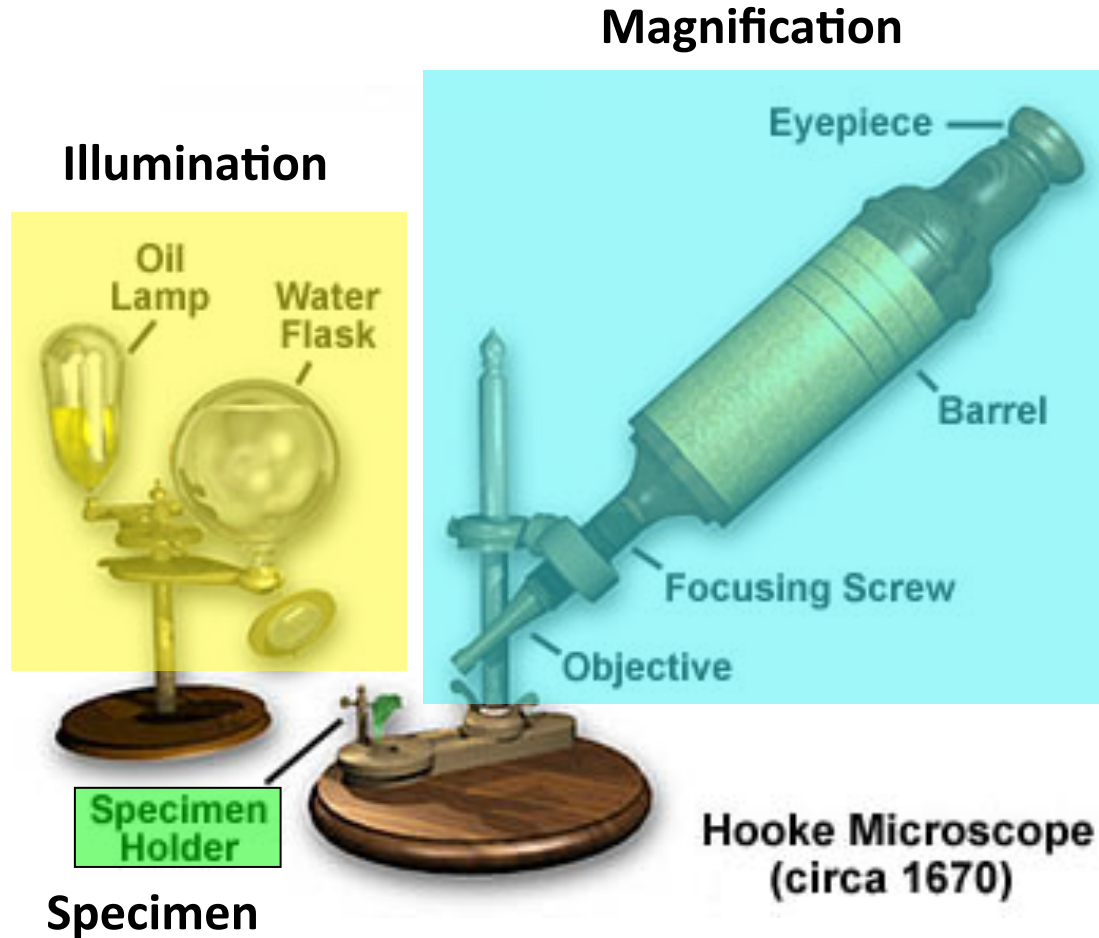
λ -wavelength

c -speed of light

Conventional Light Microscopy

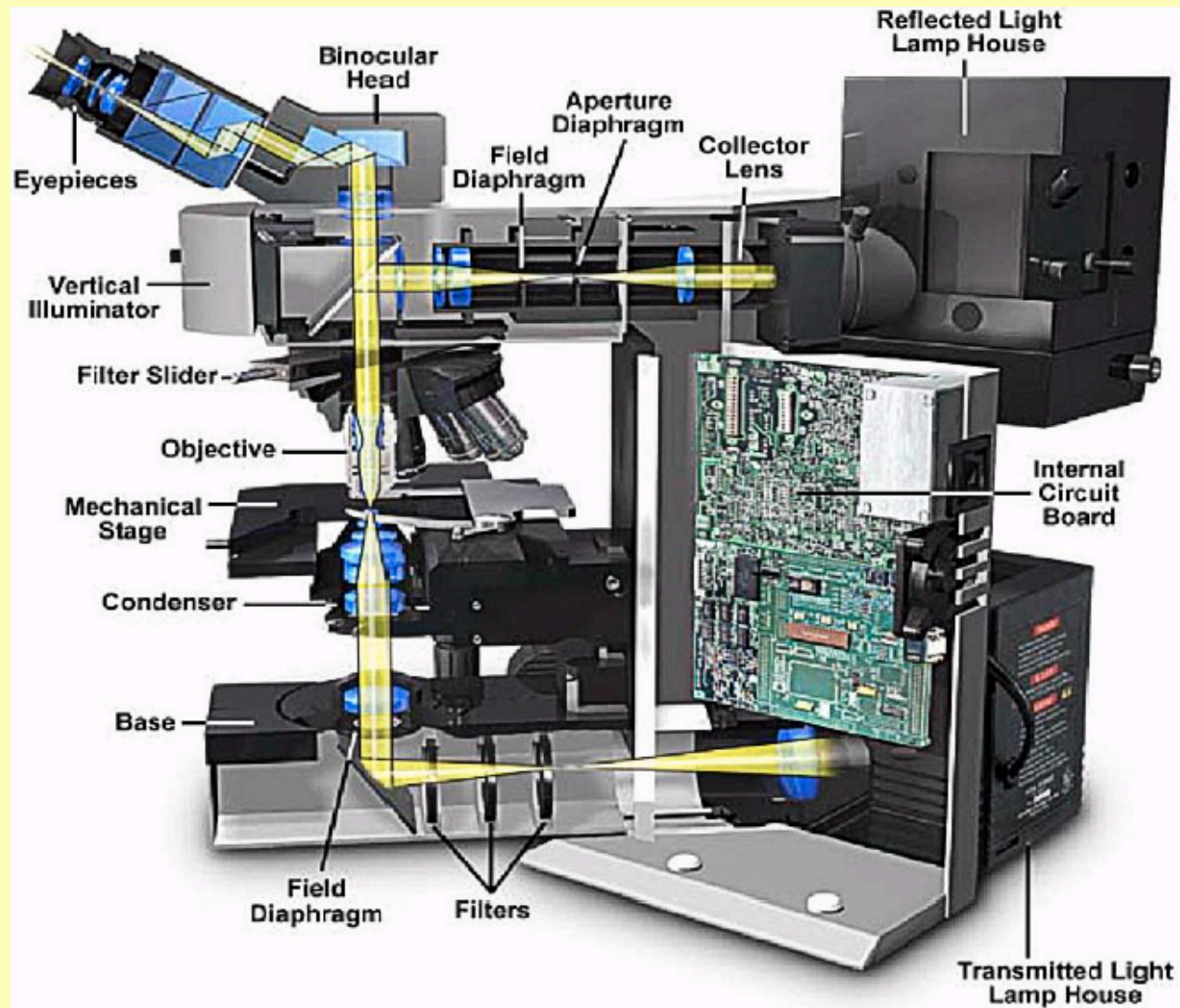
Bright field, phase & Interference contrast

Basic Anatomy of Light Microscope

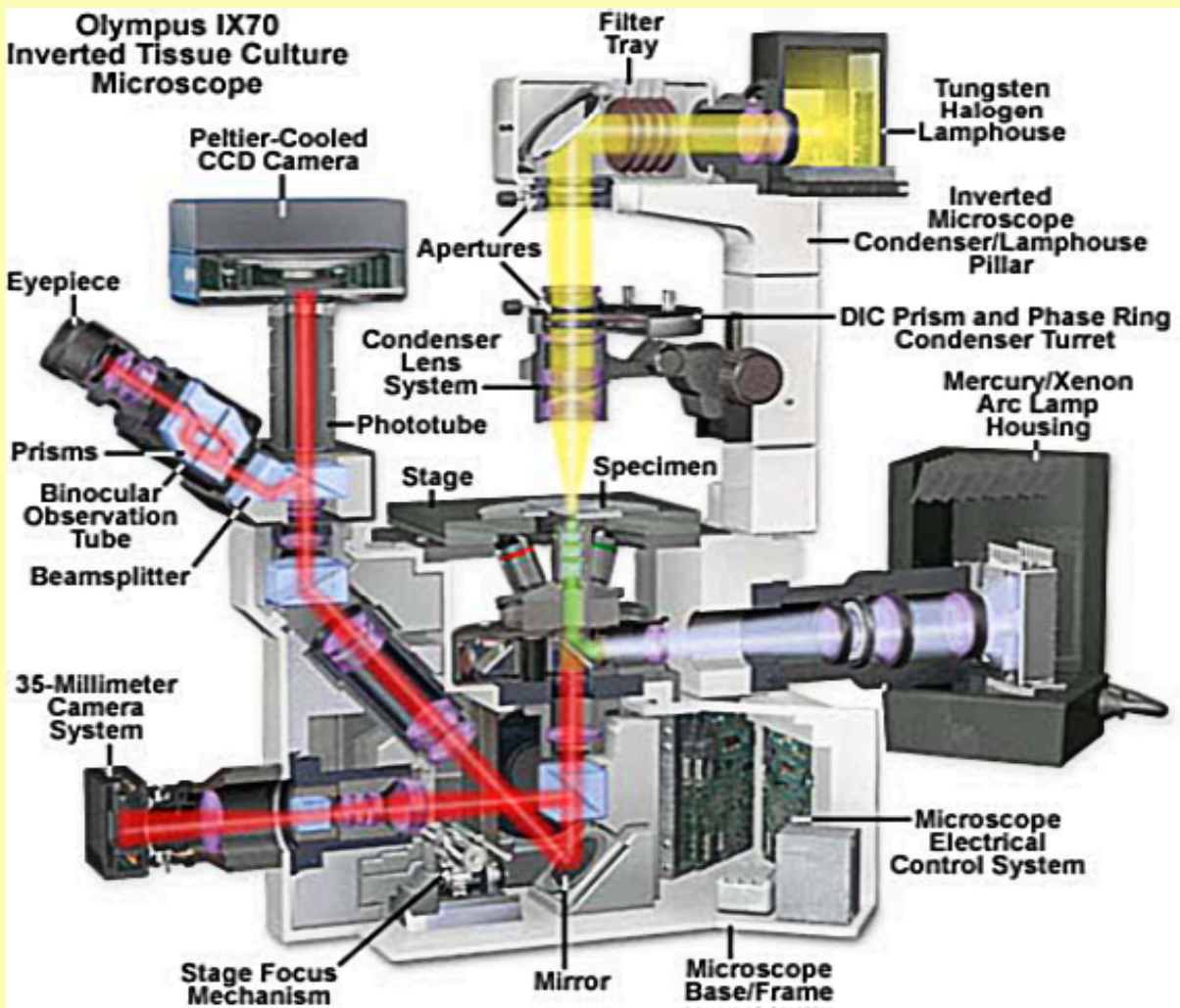


(with modification <http://micro.magnet.fsu.edu>)

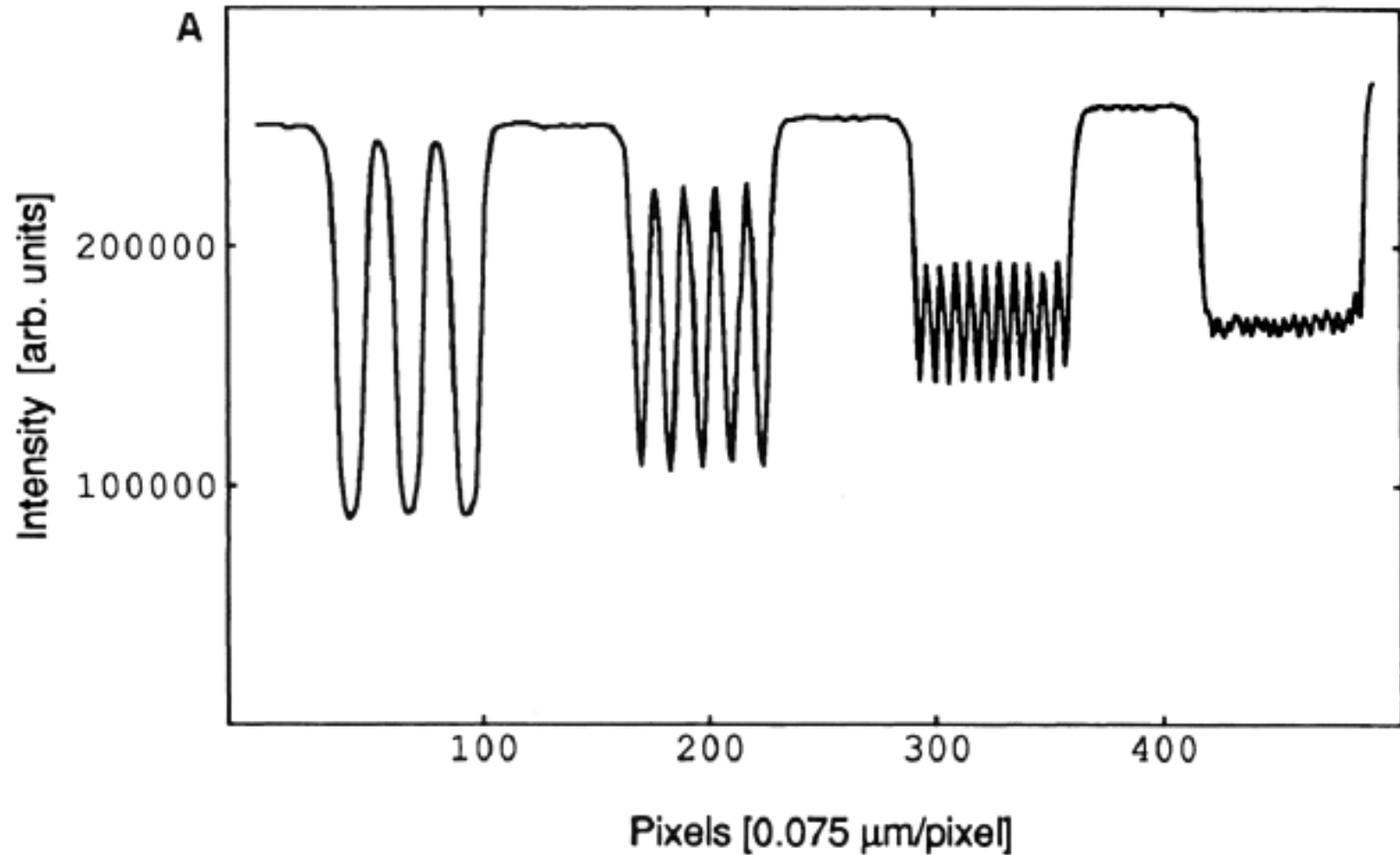
UPRIGHT MICROSCOPE



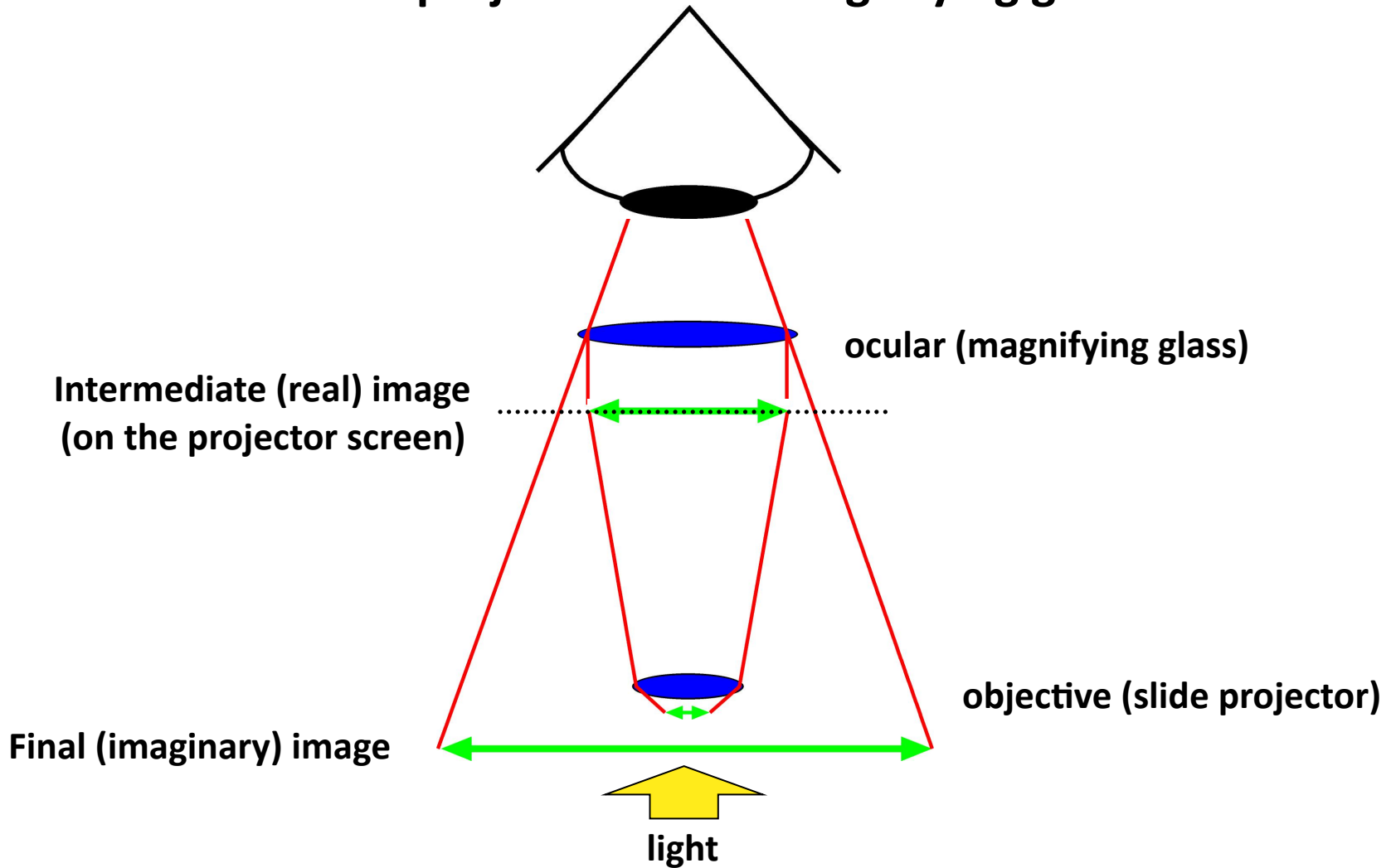
INVERTED MICROSCOPE



WHAT IS CONTRAST



**A light microscope is a combination
of a slide projector with a magnifying glass**

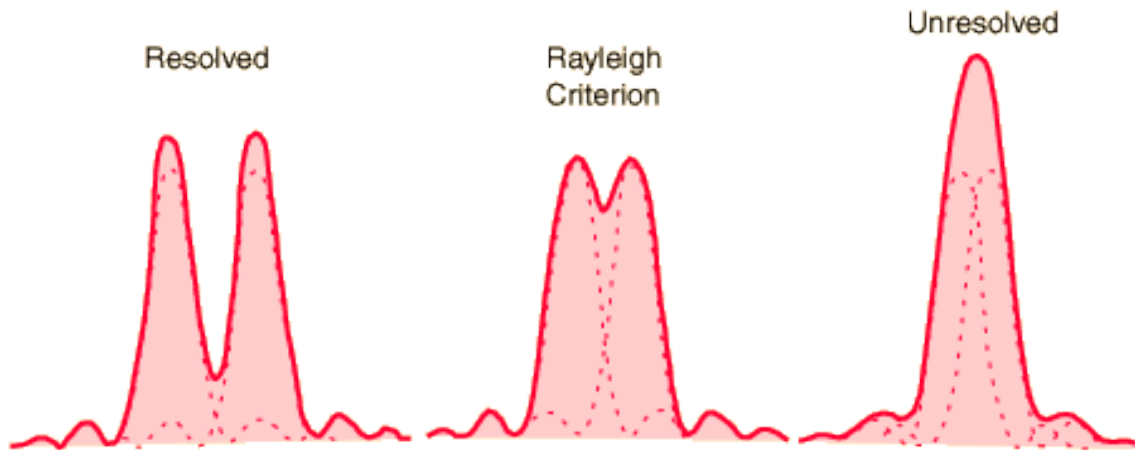


$$\text{Total magnification} = M_{\text{objective}} \times M_{\text{ocular}}$$

Rayleigh Criterion- Diffraction limit of light

Or capability of resolving adjacent objects in a microscope

P.S.F - Point Spread Function



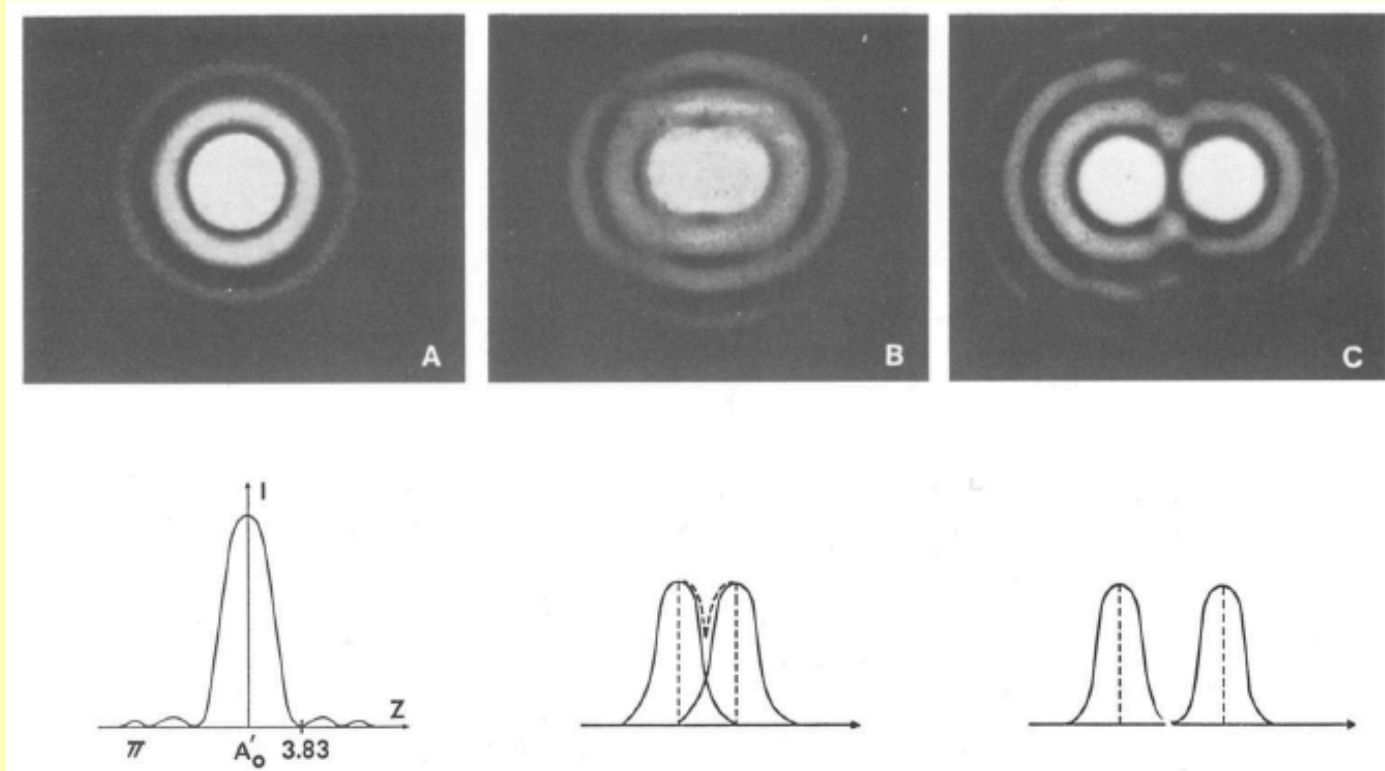
Lord Rayleigh (John Strutt)
(1842-1919)

$$R = \frac{0.61\lambda}{NA}$$

$$NA = n \sin \theta$$

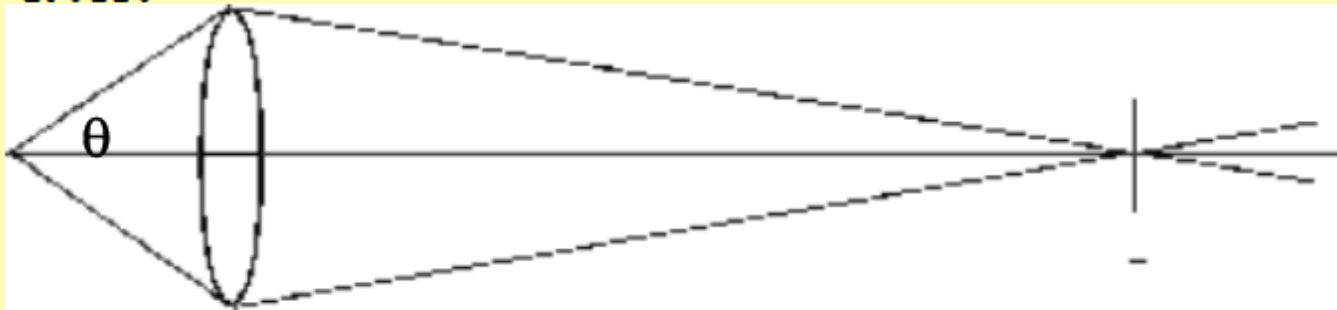
Angular collection ability of
an objective lens

DIFFRACTION LIMITED IMAGING

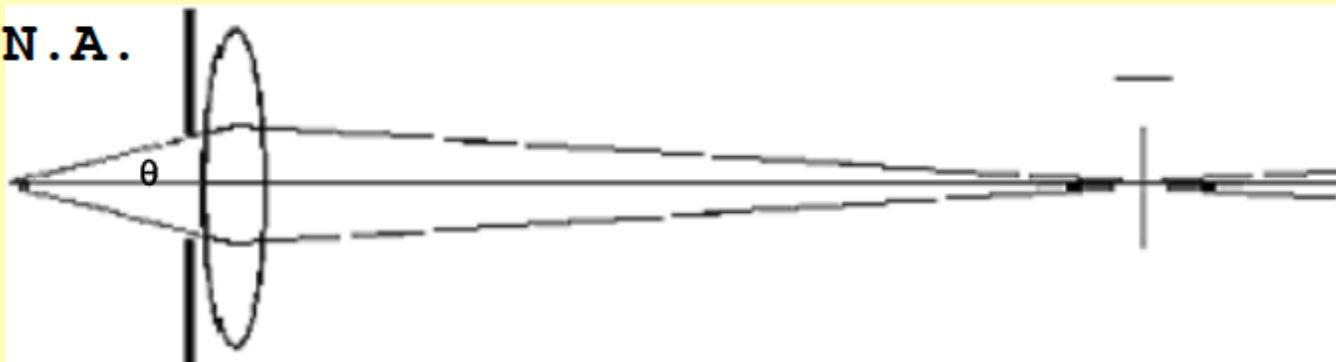


NUMERICAL APERTURE

High N.A.



Low N.A.



$$\text{N.A.} = n \sin(\theta)$$

RAYLEIGH CRITERION

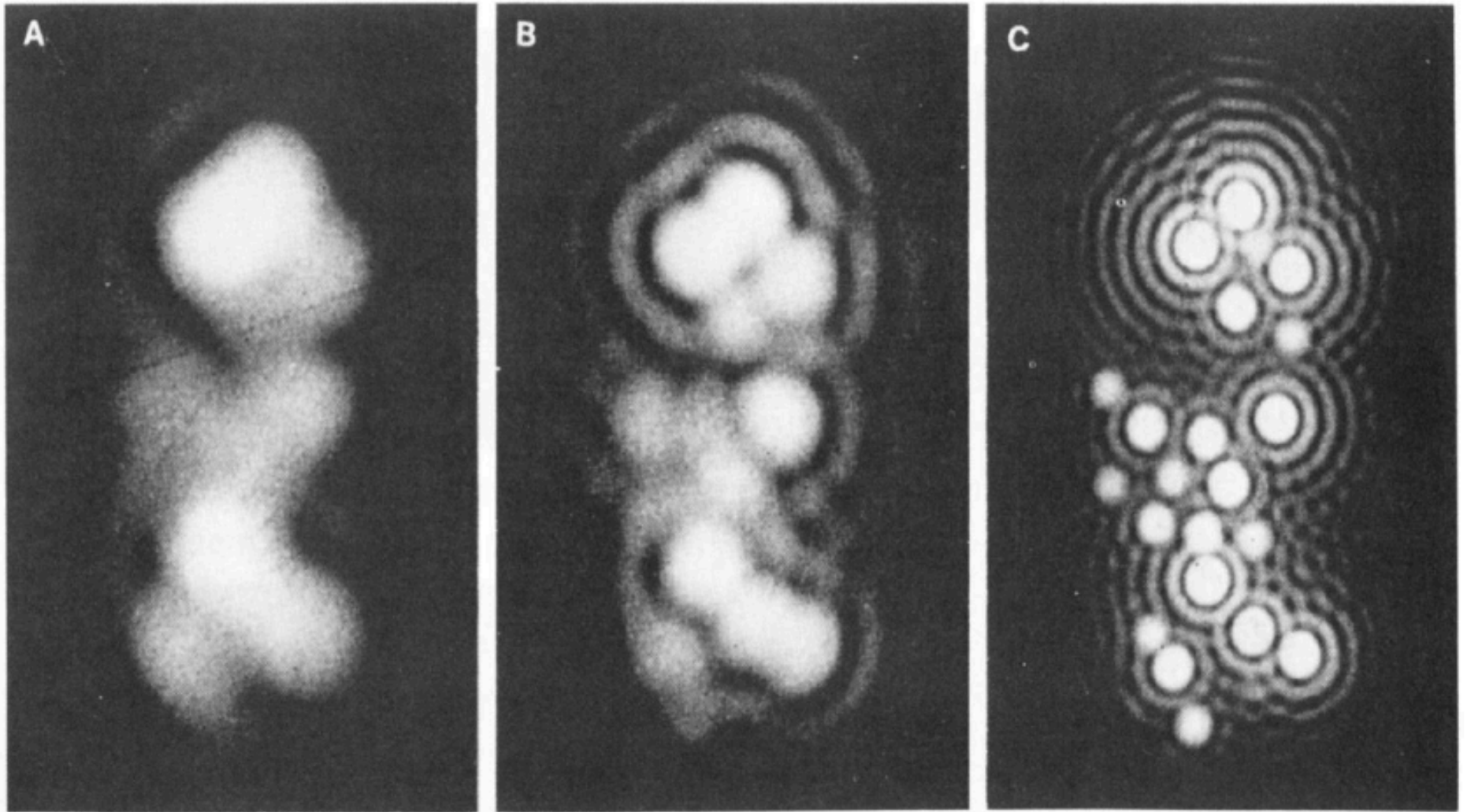
$$r = 0.61\lambda/\text{NA}$$

$$\text{NA} = n \sin(\theta)$$

For high NA objectives (0.4-1.4)

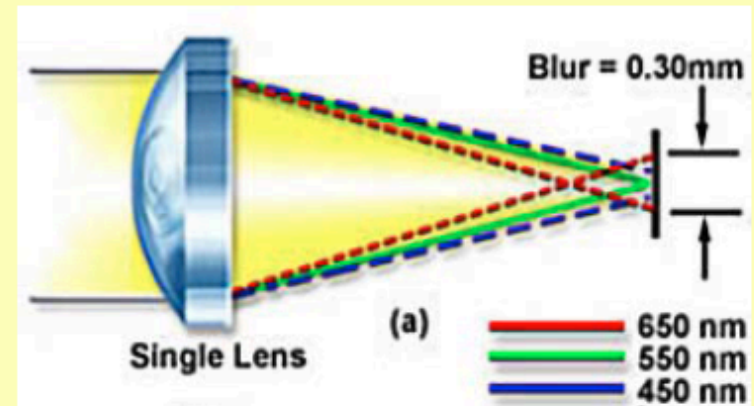
$$1.5\lambda > r > 0.44\lambda$$

EFFECT OF NA ON RESOLUTION

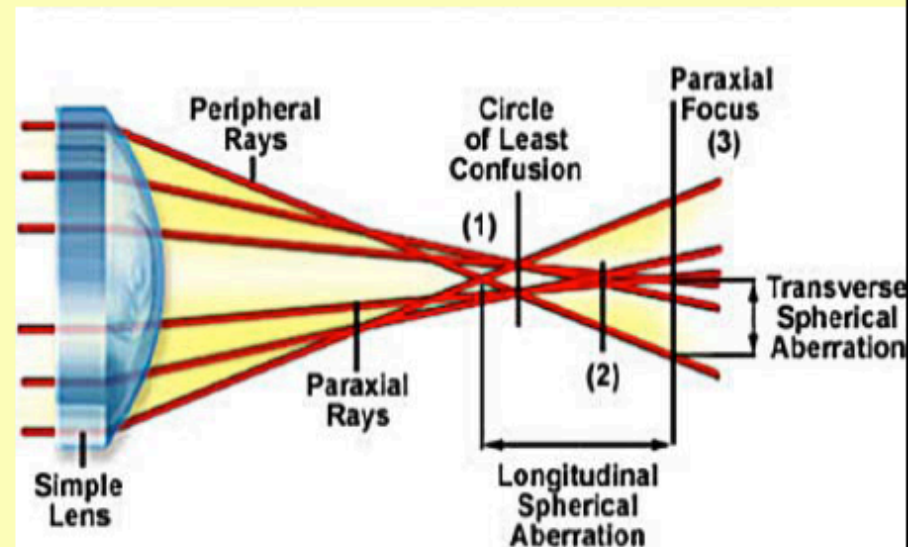


ABERRATIONS

- **CHROMATIC ABERRATION**
- due to $n(\lambda)$

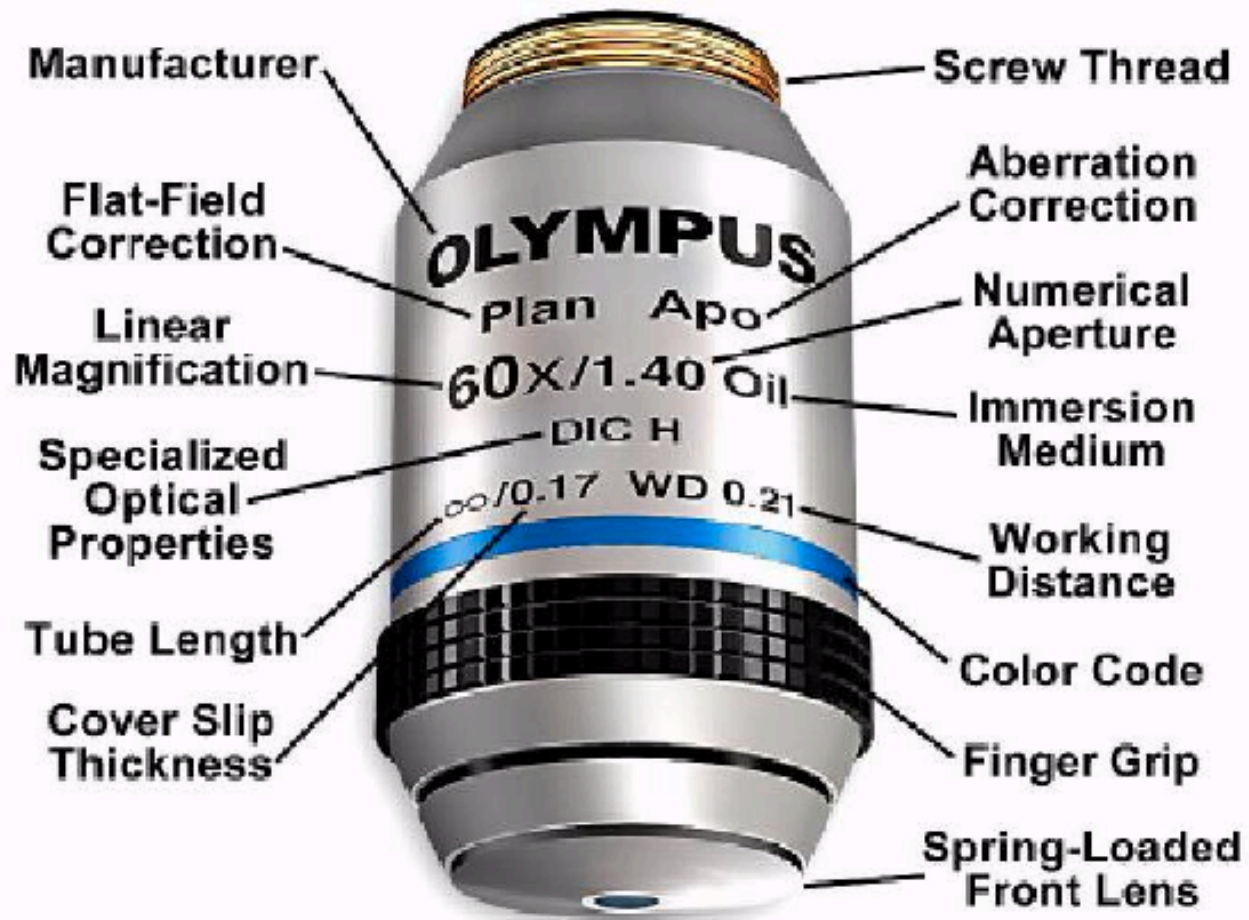


- **SPHERICAL ABERRATION**
- paraxial approx.



OBJECTIVES

Objective Specifications



WHY SO EXPENSIVE?

Optical Correction in Objectives

Achromatic Objective



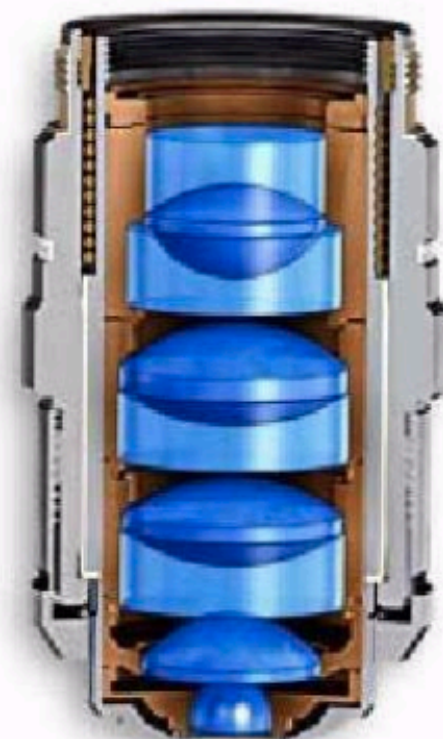
(a)

Fluorite Objective



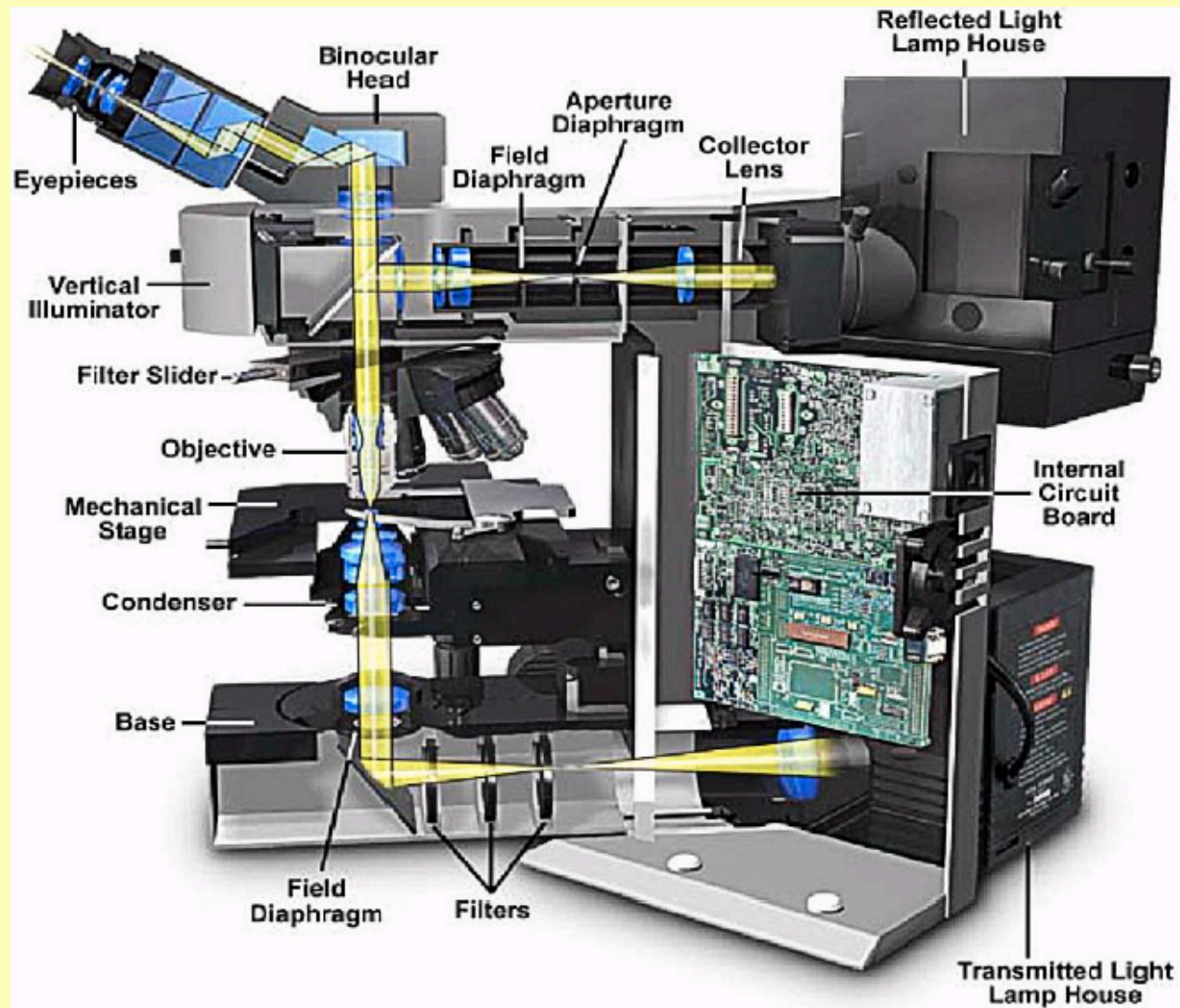
(b)

Apochromatic Objective



(c)

UPRIGHT MICROSCOPE



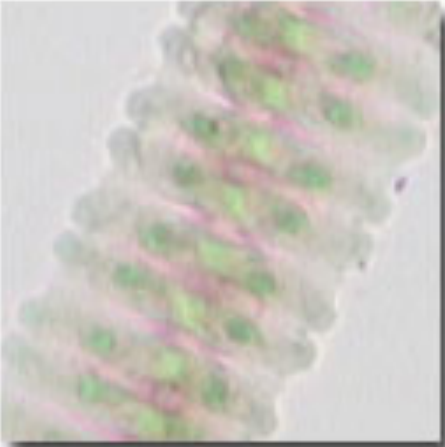
GENERATION OF CONTRAST

- Darkfield
- Rheinberg illumination
- Phase contrast microscopy
- DIC (Nomarski)

Using Properties of Light for Better Imaging

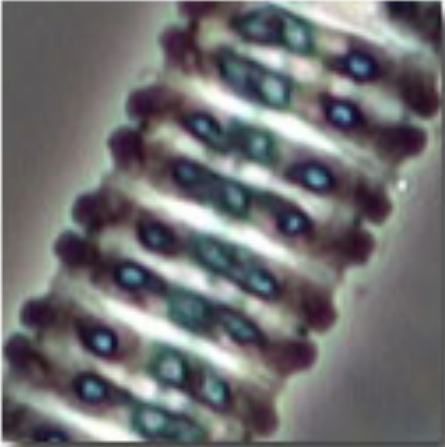
Contrast Modes in Transmitted Optical Microscopy

Algae



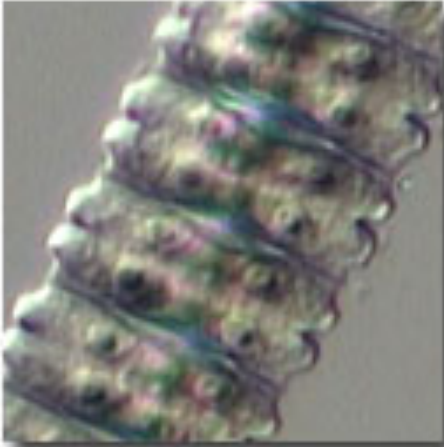
(a)

Bright field



(b)

phase

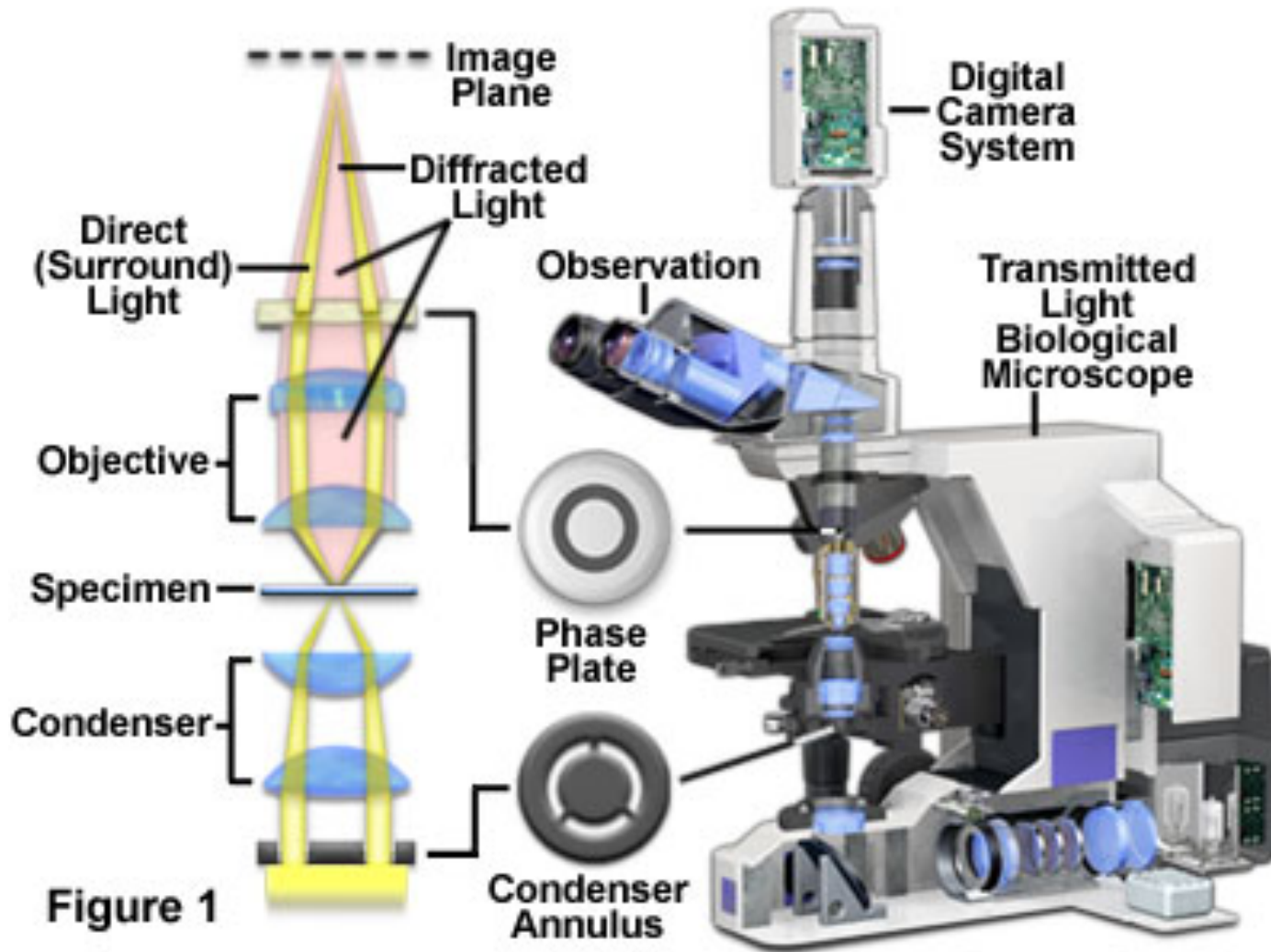


(c)

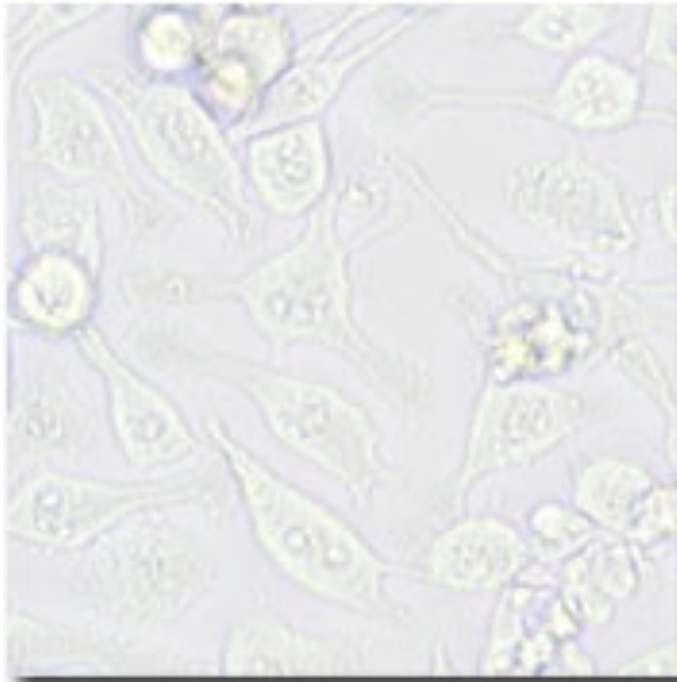
DIC

Figure 1

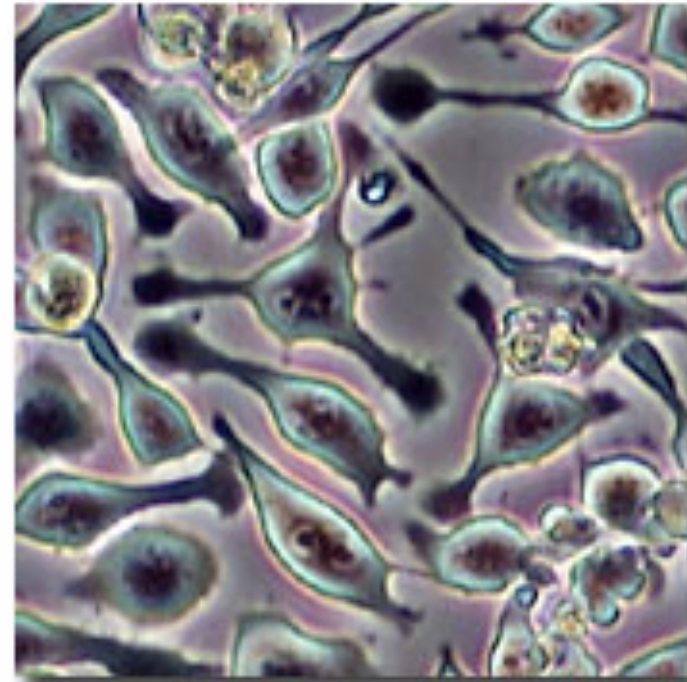
Phase Contrast Microscope Configuration



Living Cells in Brightfield and Phase Contrast



(a)



(b)

Figure 2

Phase Contrast Microscope Optical Train

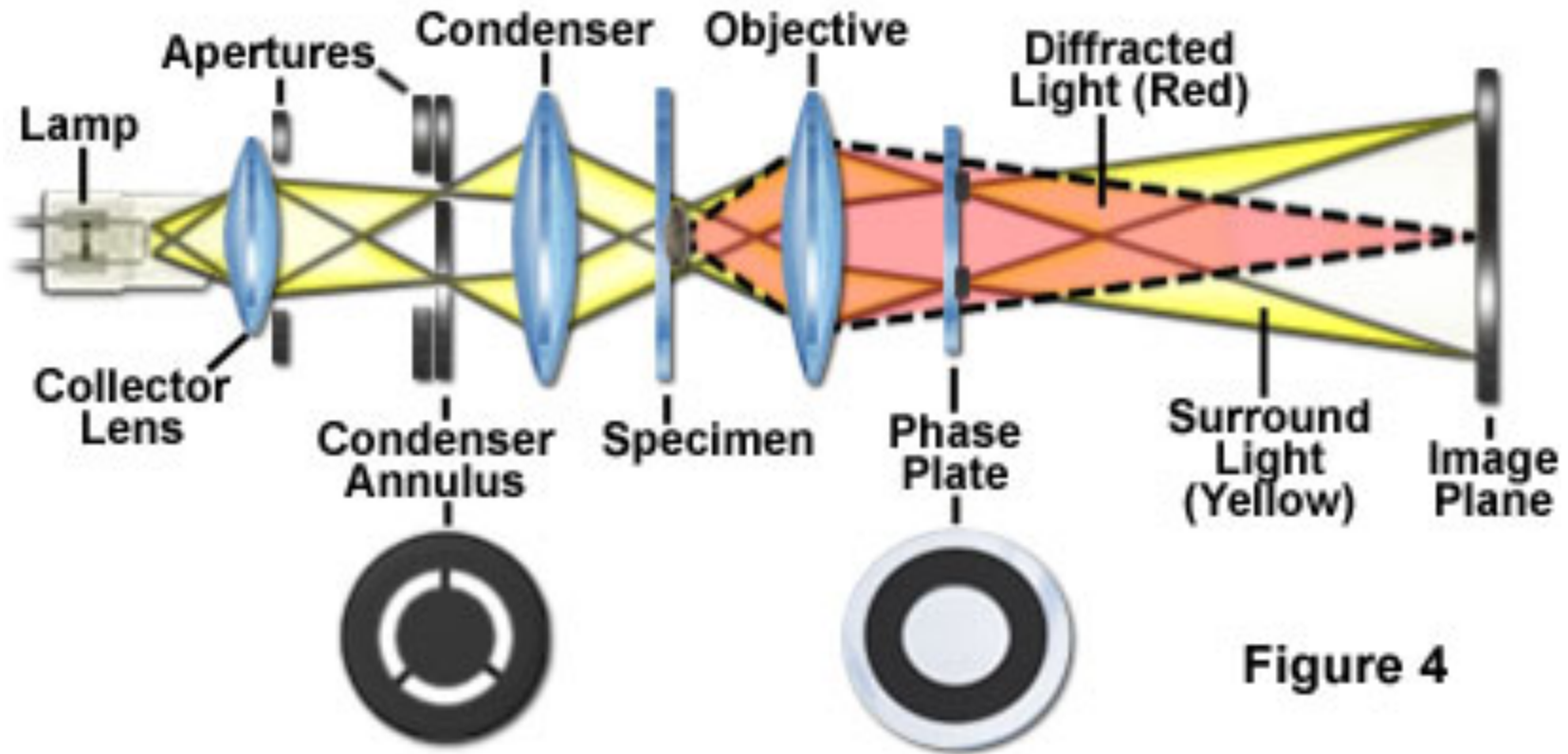
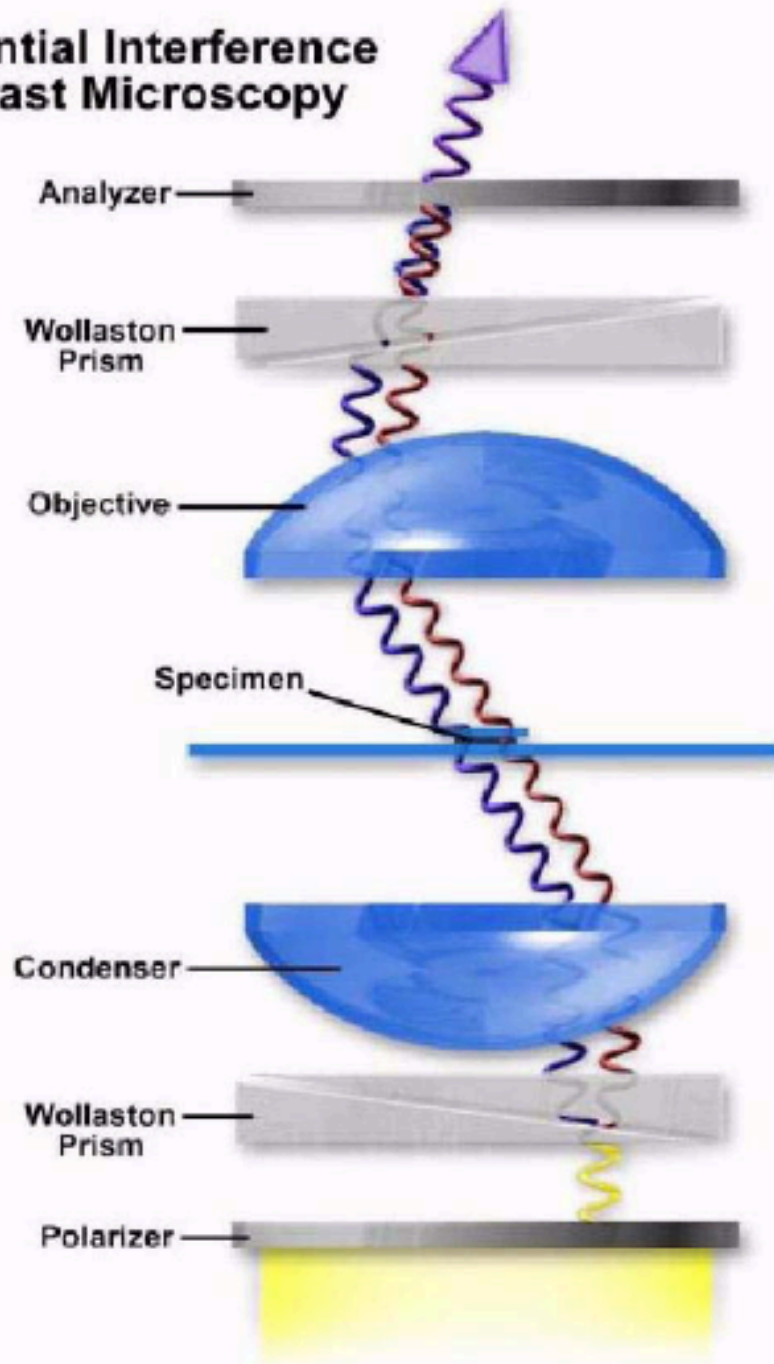
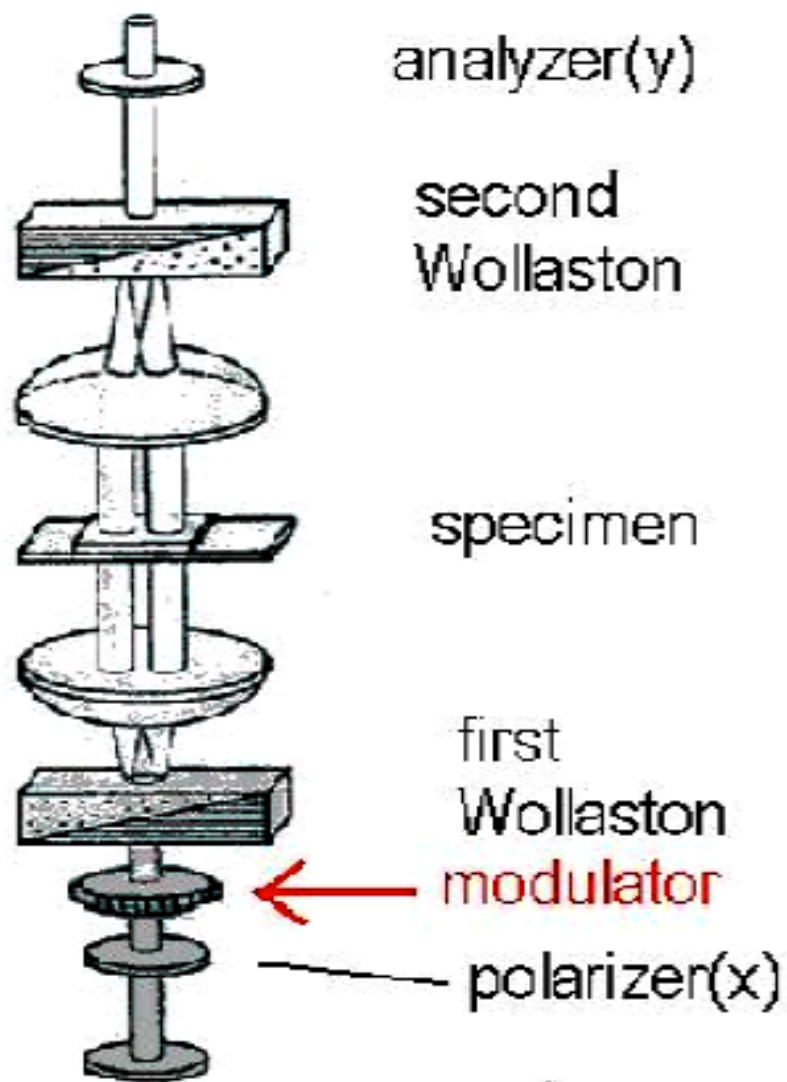


Figure 4

Differential Interference Contrast Microscopy

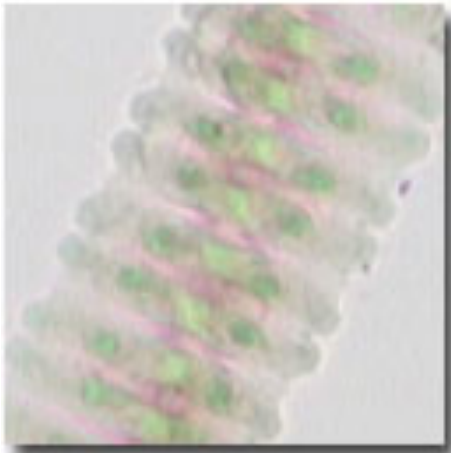




Holzwarth, Webb, Kubinski, and Allen,
J. Microscopy, p249-254 (1997)

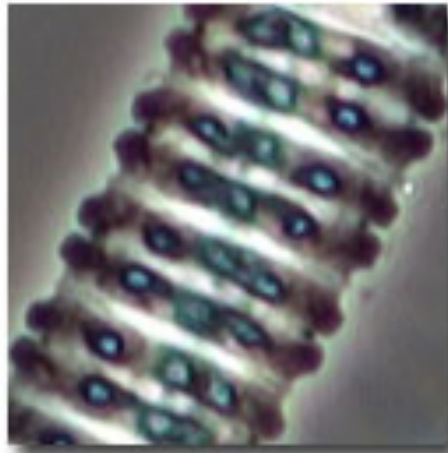
Algae

Contrast Modes in Transmitted Optical Microscopy



(a)

Bright field



(b)

phase

Figure 1

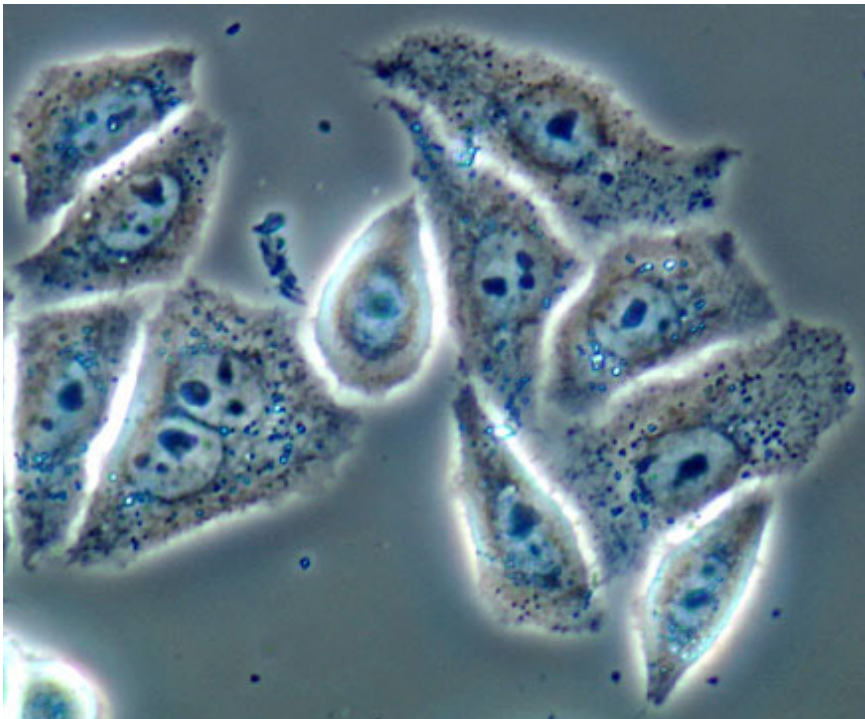


(c)

DIC

Phase vs DIC

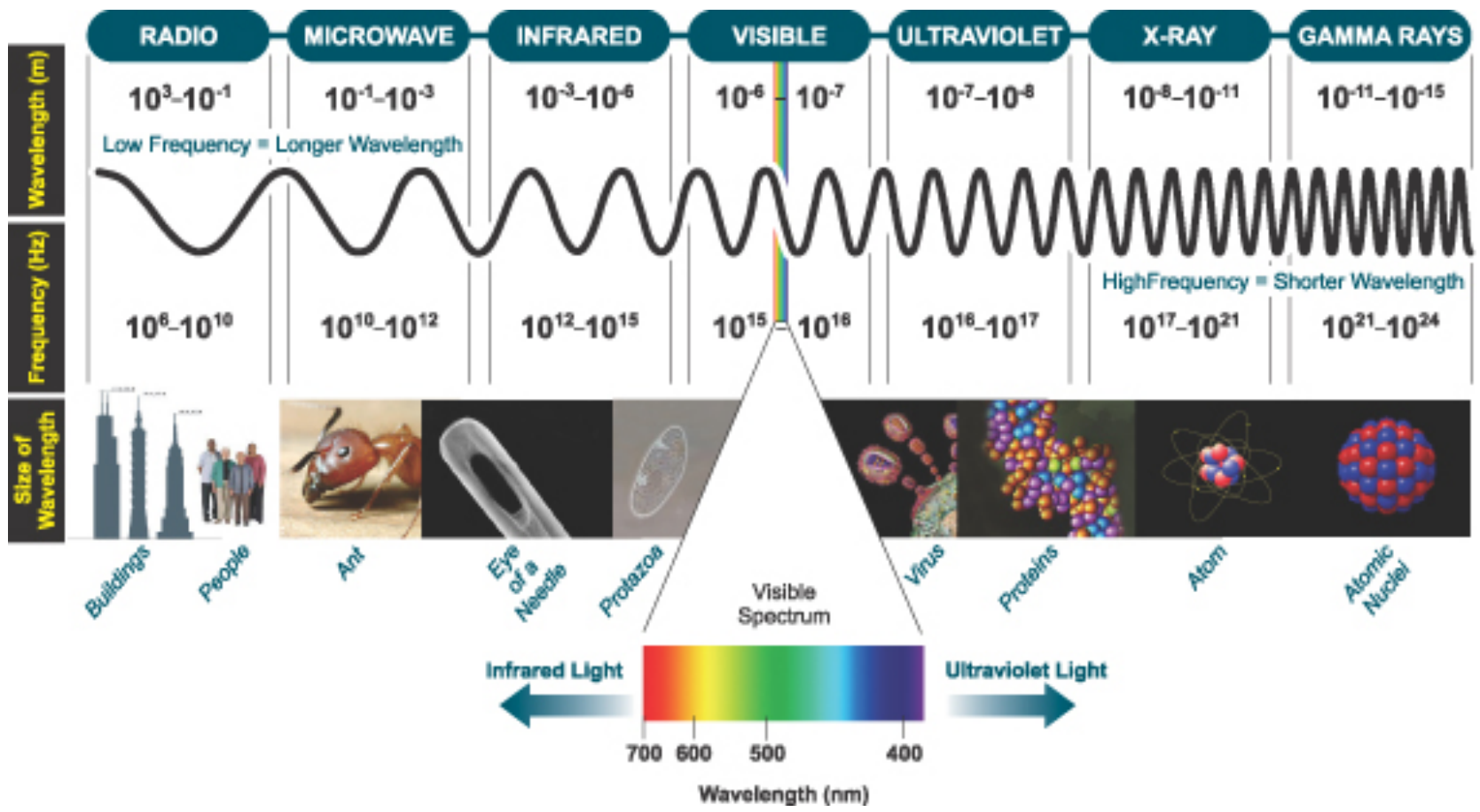
HeLa Cells



phase

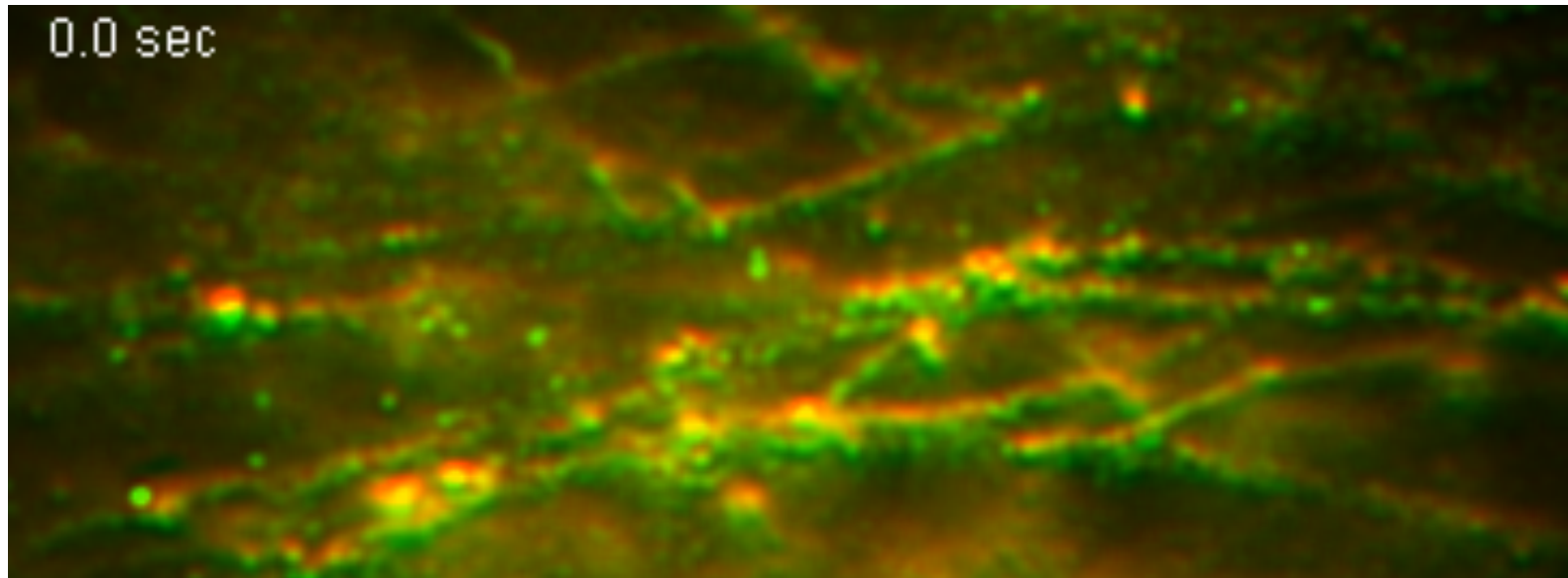


DIC



Andor Technology

Fluorescence Microscopy



Rat Neuron

Why Fluorescence Microscopy

Cells ~ 40-100 μm (10^{-6} m)

Inside the cell:

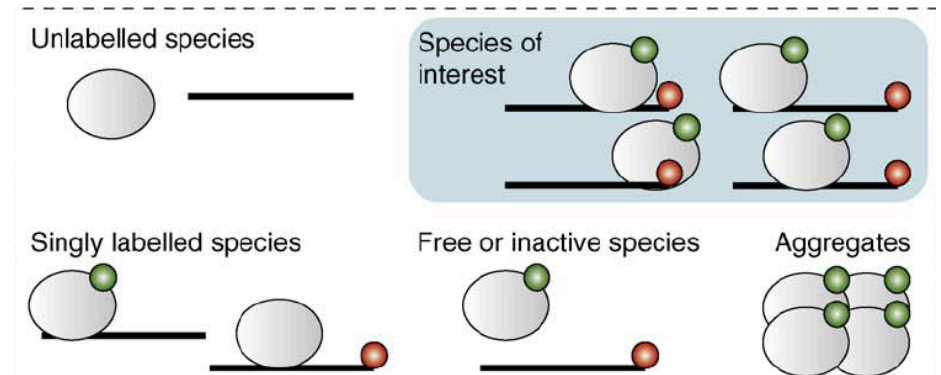
Proteins and nucleic acids – nanometer and angstrom scale (10^{-9} - 10^{-10} m).

Seeing biology as it happens – light microscopy (Visualizing Small Objects)

Problem: Small objects will not scatter light

Solution: Fluorescence microscopy

Instead of looking directly we attach to the biomolecules of interest a fluorescent molecule (Tag)



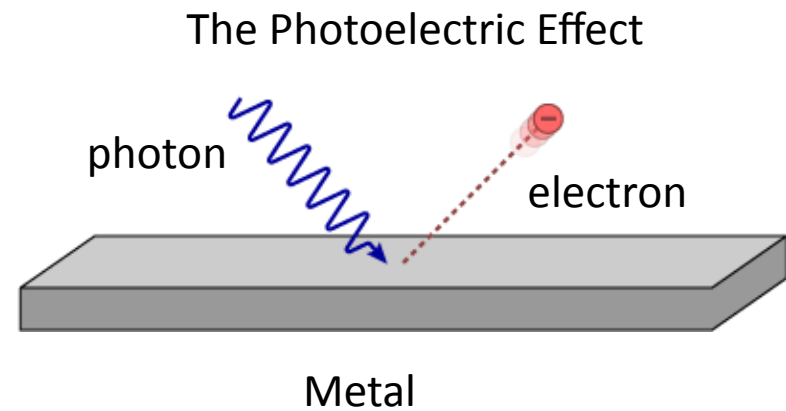
Fluorescent Molecules:

- **Fluorophores/Fluorochromes/Dye molecules**
- **Fluorescent Proteins (Bioluminescence)**
- **Quantum-Dots**

The Duality of Light: **What is the Physical Nature of Light?**

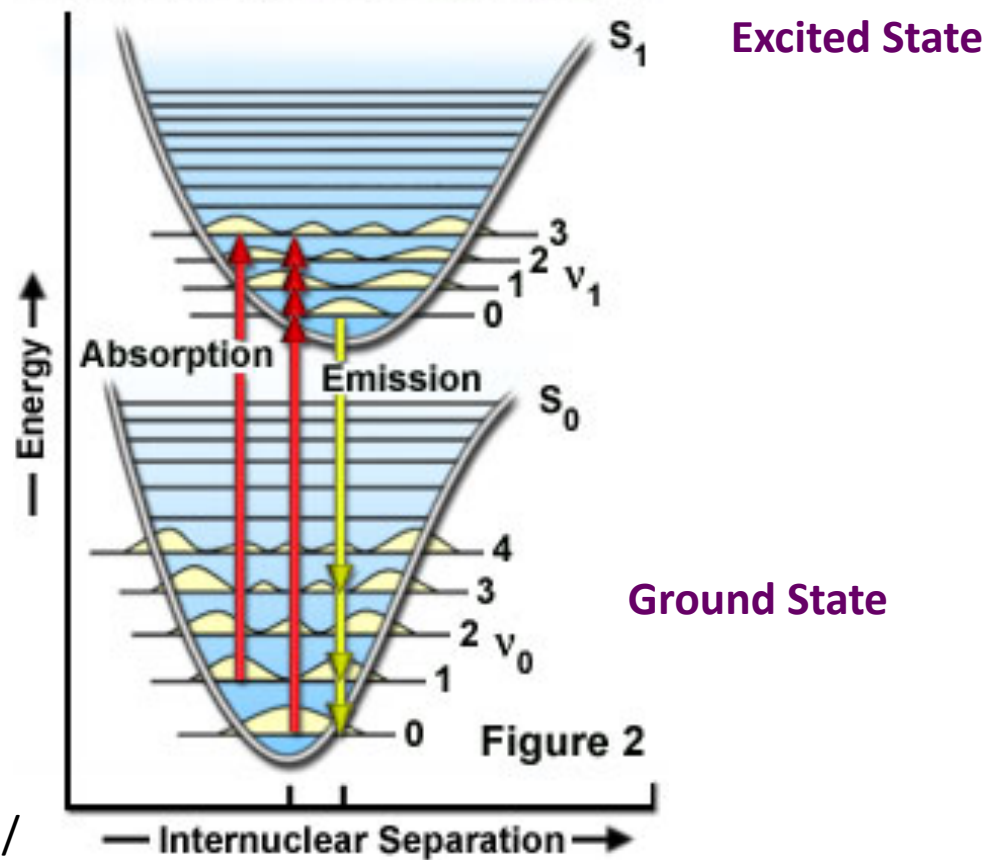
Light behaves like.... Particles
(Discrete Quantities of Energy- light quantas or photons)

Hertz, Thomson, Lenard, Einstein

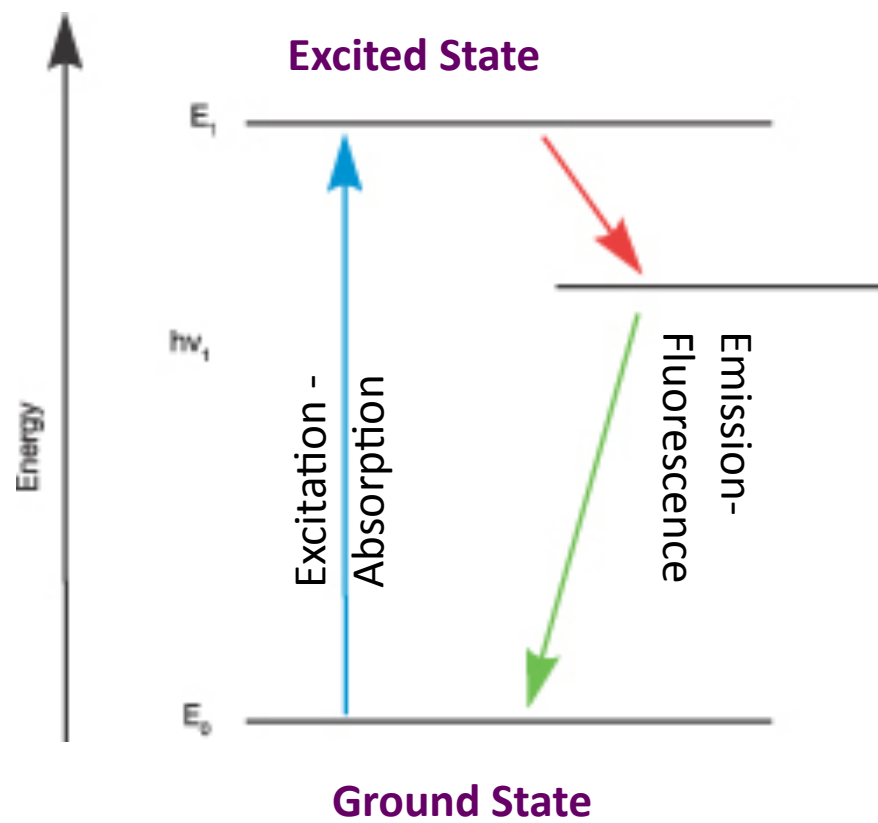


Fluorescence

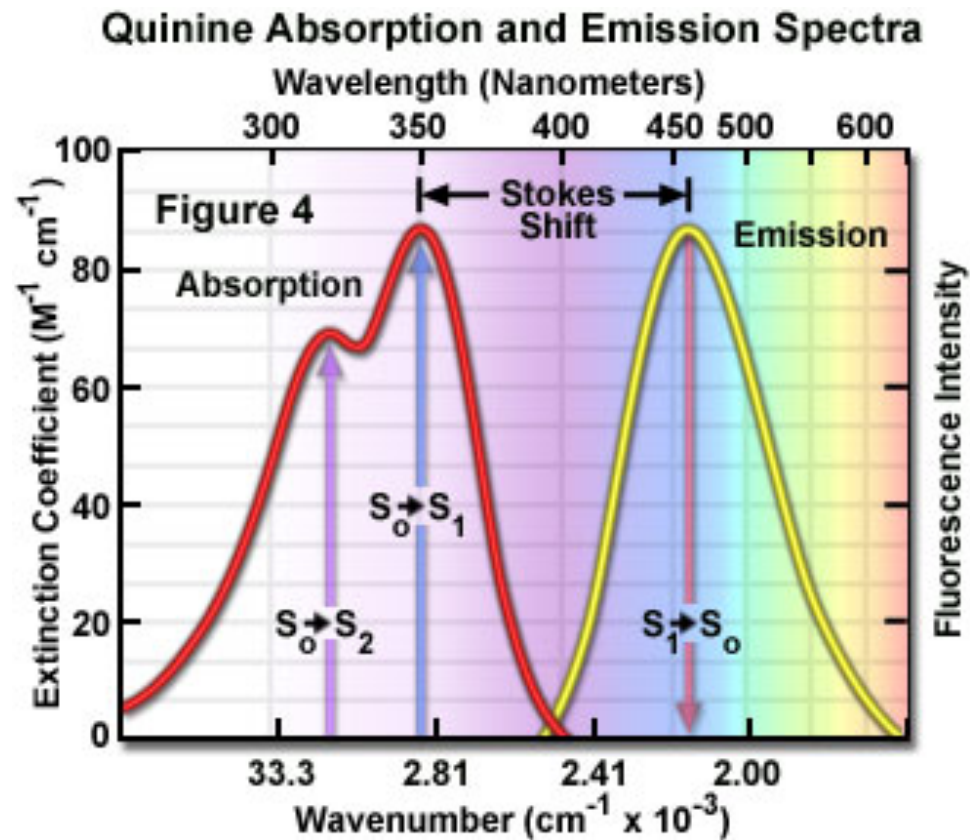
Franck-Condon Energy Diagram



Fluorescence



Fluorescence



Stokes Shift – Fluorescence Microscopy

Fluorescence

Absorption/Excitation



Fluorescence



Energy Transfer



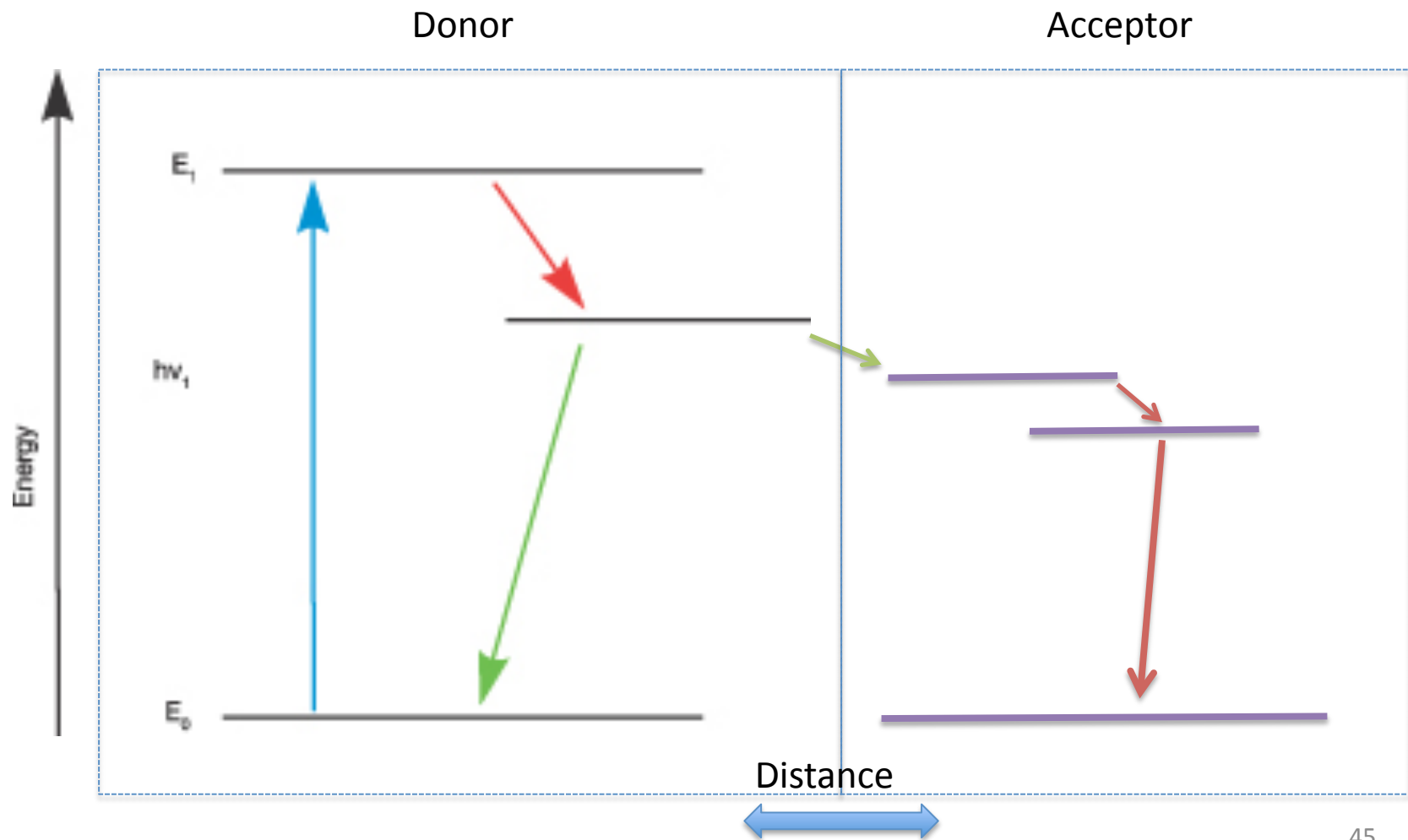
Electron Transfer



Non-Radiative Decay



Fluorescence Resonance Energy Transfer (FRET)



Fluorescence Resonance Energy Transfer (FRET) “Spectroscopic Ruler”

$$2 \text{ nm} < R < 8 \text{ nm}$$

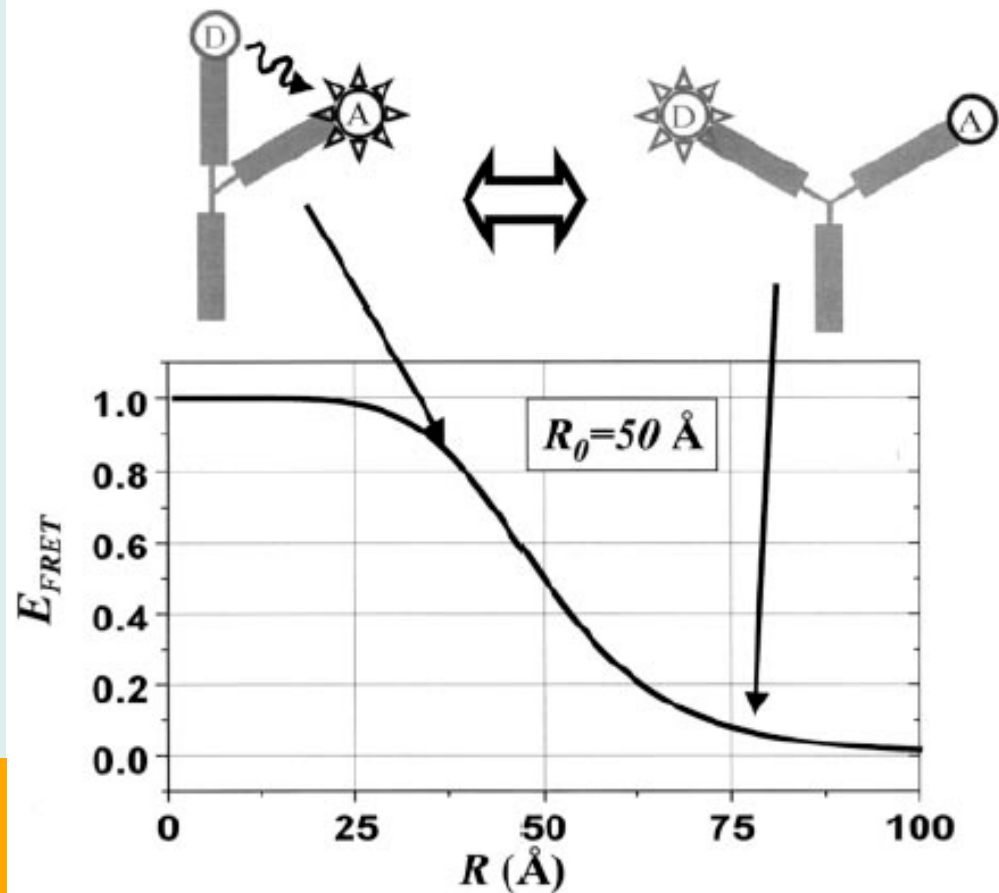
$$E_{\text{FRET}} = I_A / (I_A + I_D)$$

$$E_{\text{FRET}} = (1 + (R/R_0)^6)^{-1}$$

$$R/R_0 = ((1/E_{\text{FRET}}) - 1)^{1/6}$$

$$R_0(\text{Cy3-Cy5}) = 6.3 \text{ nm}$$

MEASURE DISTANCE CHANGES OVER SMALL SCALES



Increased Acceptor Signal = Decreased Donor Signal = High Transfer (FRET) Efficiency = Shorter Distance

Fluorescent Molecules:

- **Fluorophores/Fluorochromes/Dye molecules** (Alexa, Atto, CyDye, YO-PRO etc.)

Advantages: Huge selection, small, specific, bright.

Disadvantages: Introduction into cells.

- **Fluorescent Proteins** (Bioluminescence)

Advantages: genetically encoded, growing selection, specificity.

Disadvantages: Stability, size, brightness.

- **Quantum-Dots**

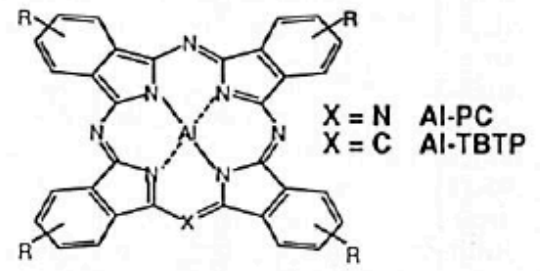
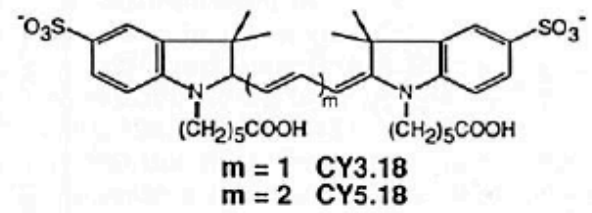
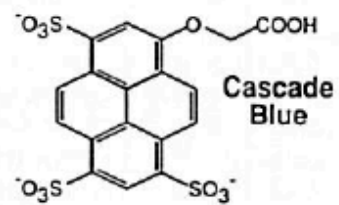
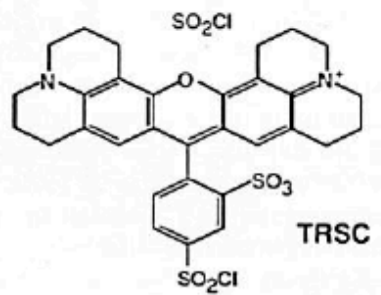
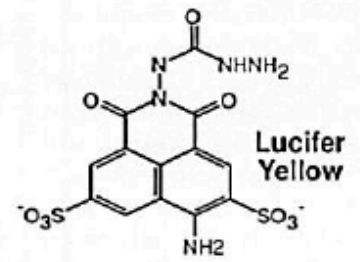
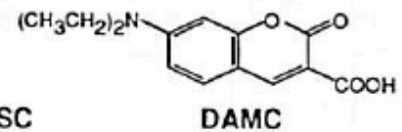
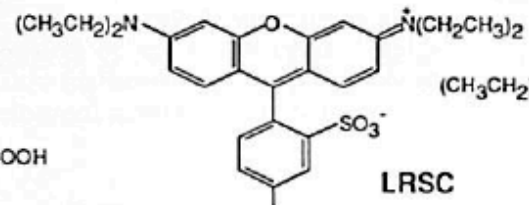
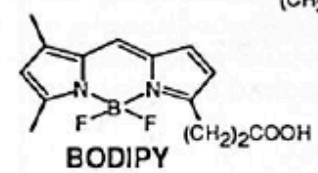
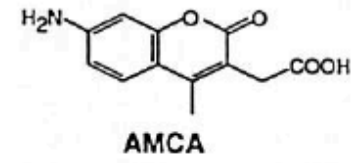
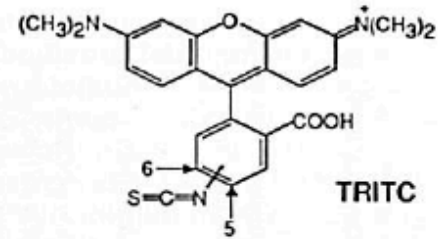
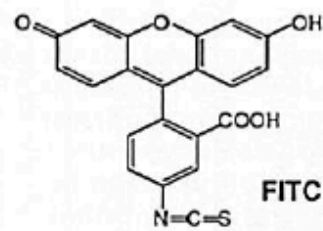
Advantages: extremely bright and stable, wide selection, continuous excitation.

Disadvantages: Very large, hard to conjugate.

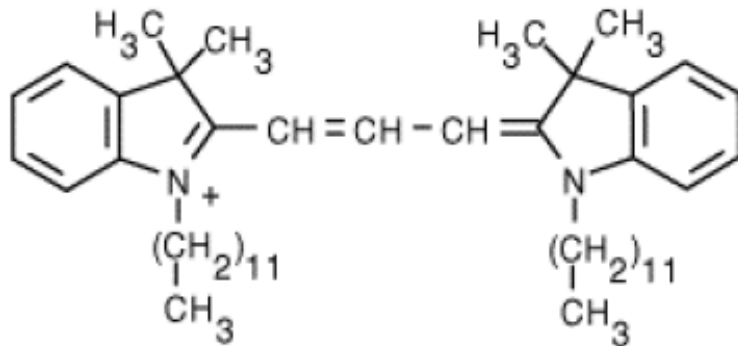
TWO IMPORTANT PROPERTIES

- Quantum efficiency:
photons emitted/photons absorbed
- Photobleaching efficiency:
probability of bleaching/photon absorbed

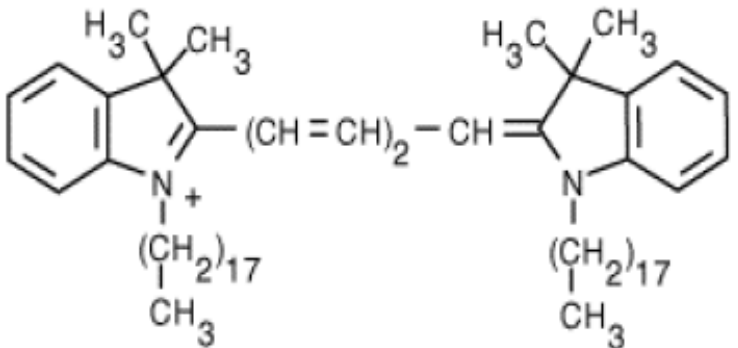
SOME COMMON FLUOROPHORES



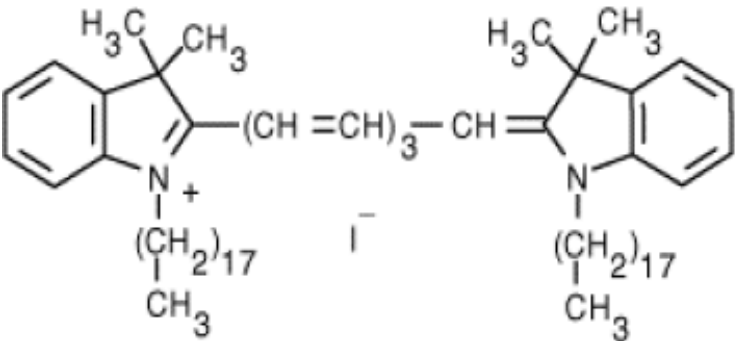
DYES COME IN FAMILIES



Abs = 549nm, Em = 565nm



Abs = 650nm, Em = 670nm



Abs = 748nm, Em = 780nm

THINGS DYES CAN SENSE

- Ca, Mg, Na, pH
- DNA, RNA, together or separately
- Potential (fast and slow dyes)
- Lipid vs aqueous surroundings
- Temperature
- Viscosity
- Each other

Fluorescent Proteins

Tagging with GFP

The available fluorescent proteins:

BFP

ECFP

EGFP

PAGFP

EYFP & related

dsRED

DHRed

Other choices

Where to tag?

N-terminal

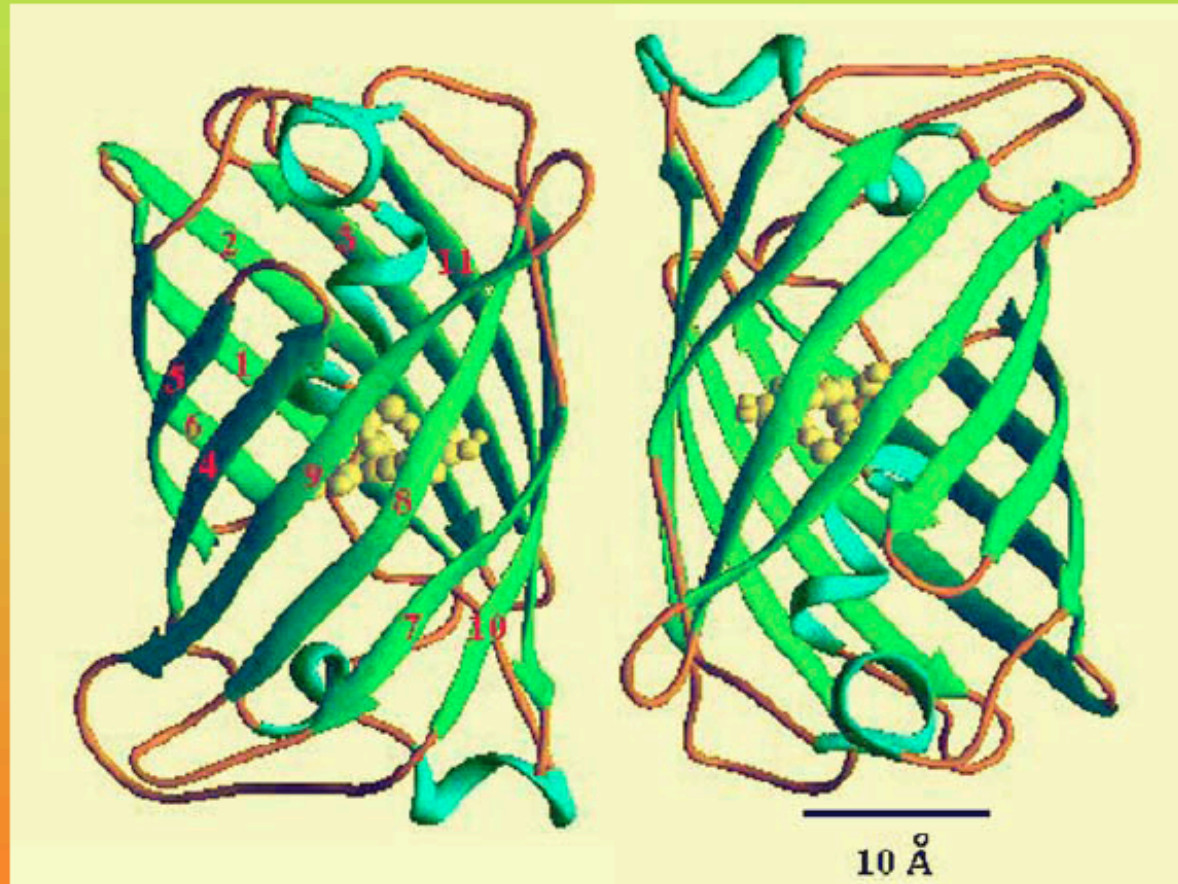
C-terminal

Localization signals

Other considerations:

Localization

function



Fluorescent Proteins

Tagging with GFP

The available fluorescent proteins:

BFP

ECFP

EGFP

PAGFP

EYFP & related dsRED

DHCred

Other choices

Where to tag?

N-terminal

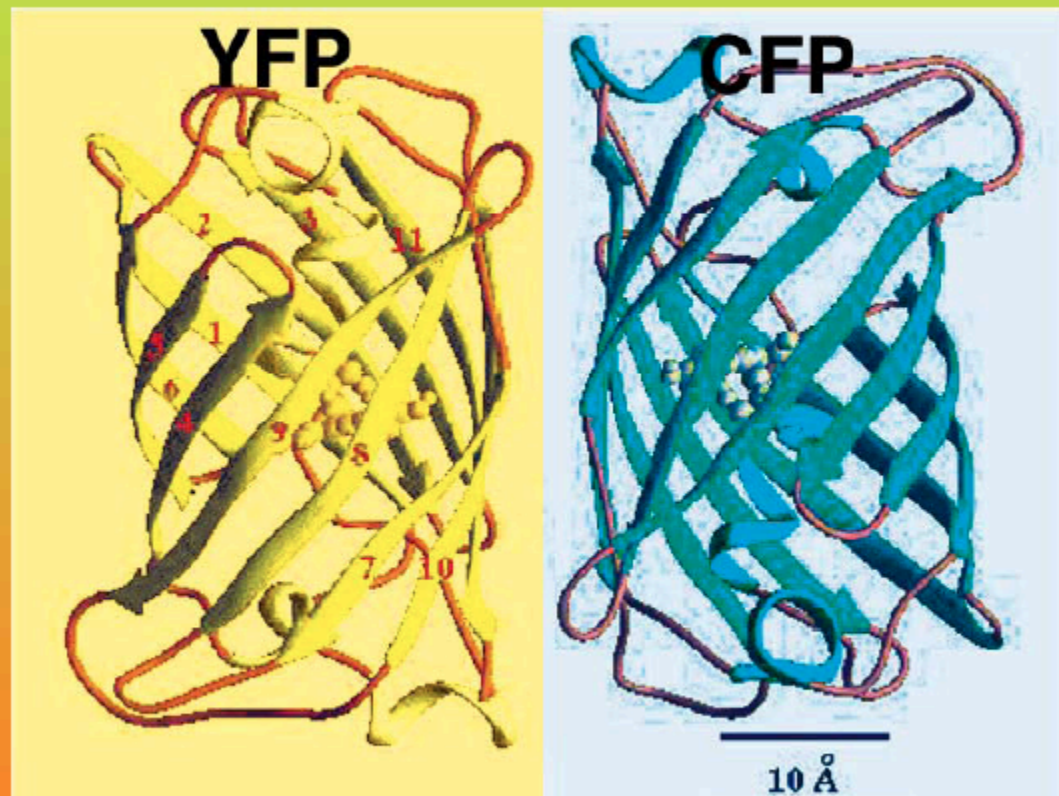
C-terminal

Localization signals

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Fluorescent Proteins

Tagging with GFP

The available fluorescent proteins:

BFP

ECFP

EGFP

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EYFP & related

dsRED

DHCRred

Other choices

Where to tag?

N-terminal

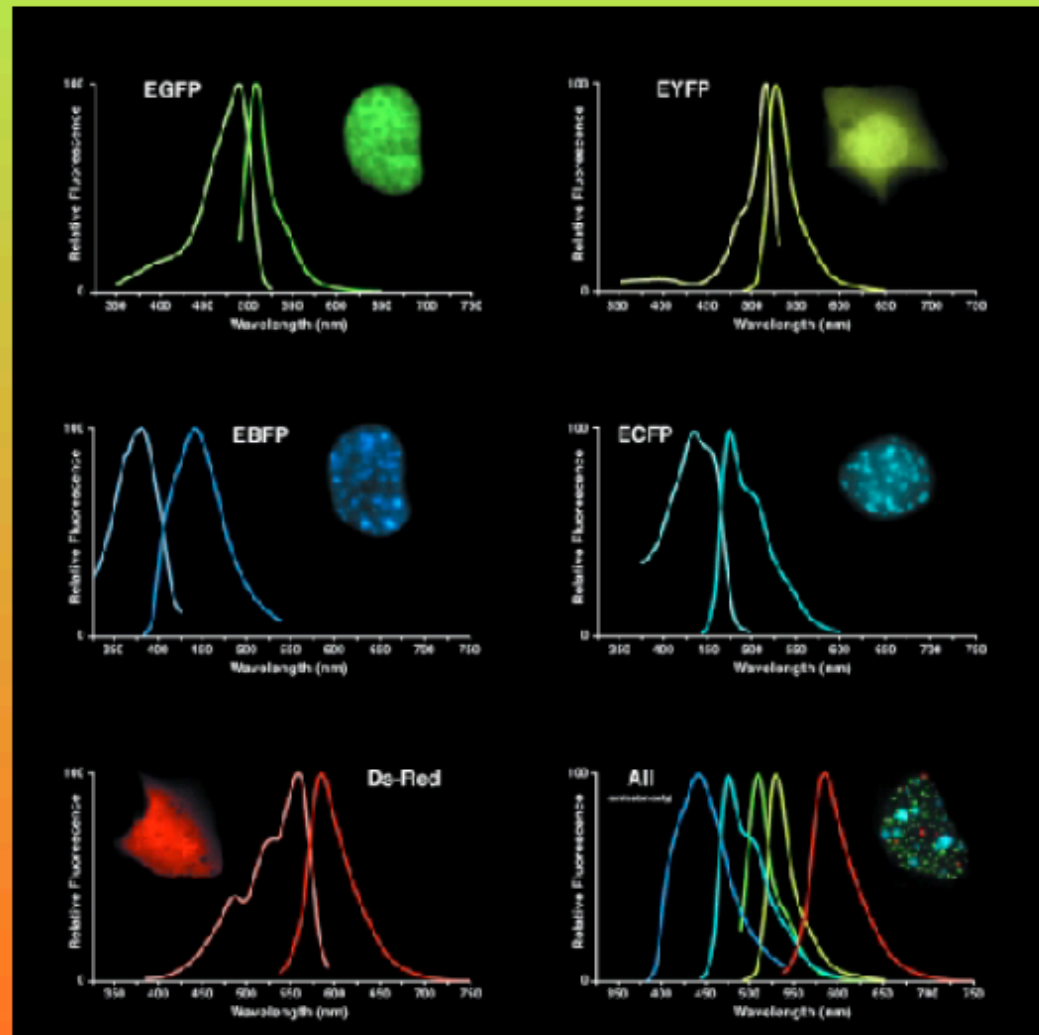
C-terminal

Localization signals

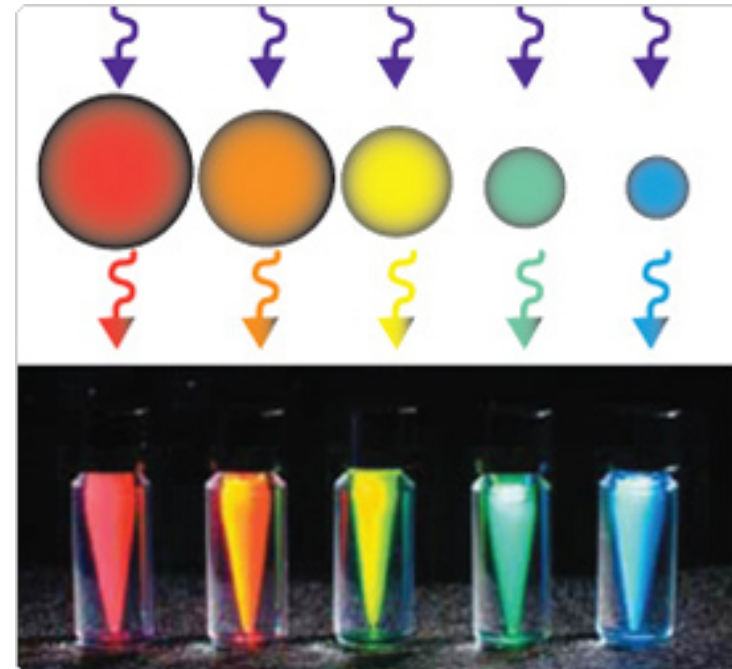
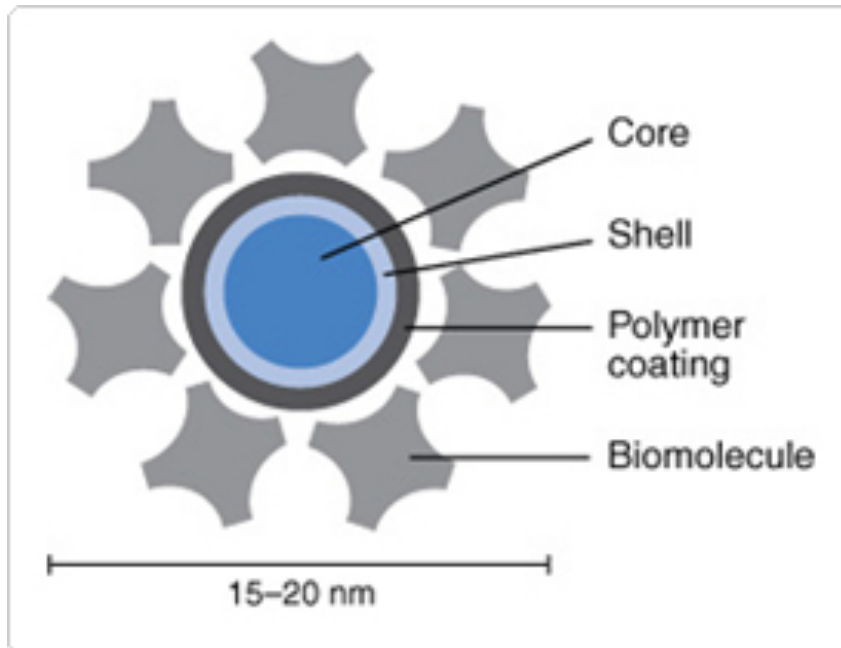
Other considerations:

Localization

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Quantum-Dots

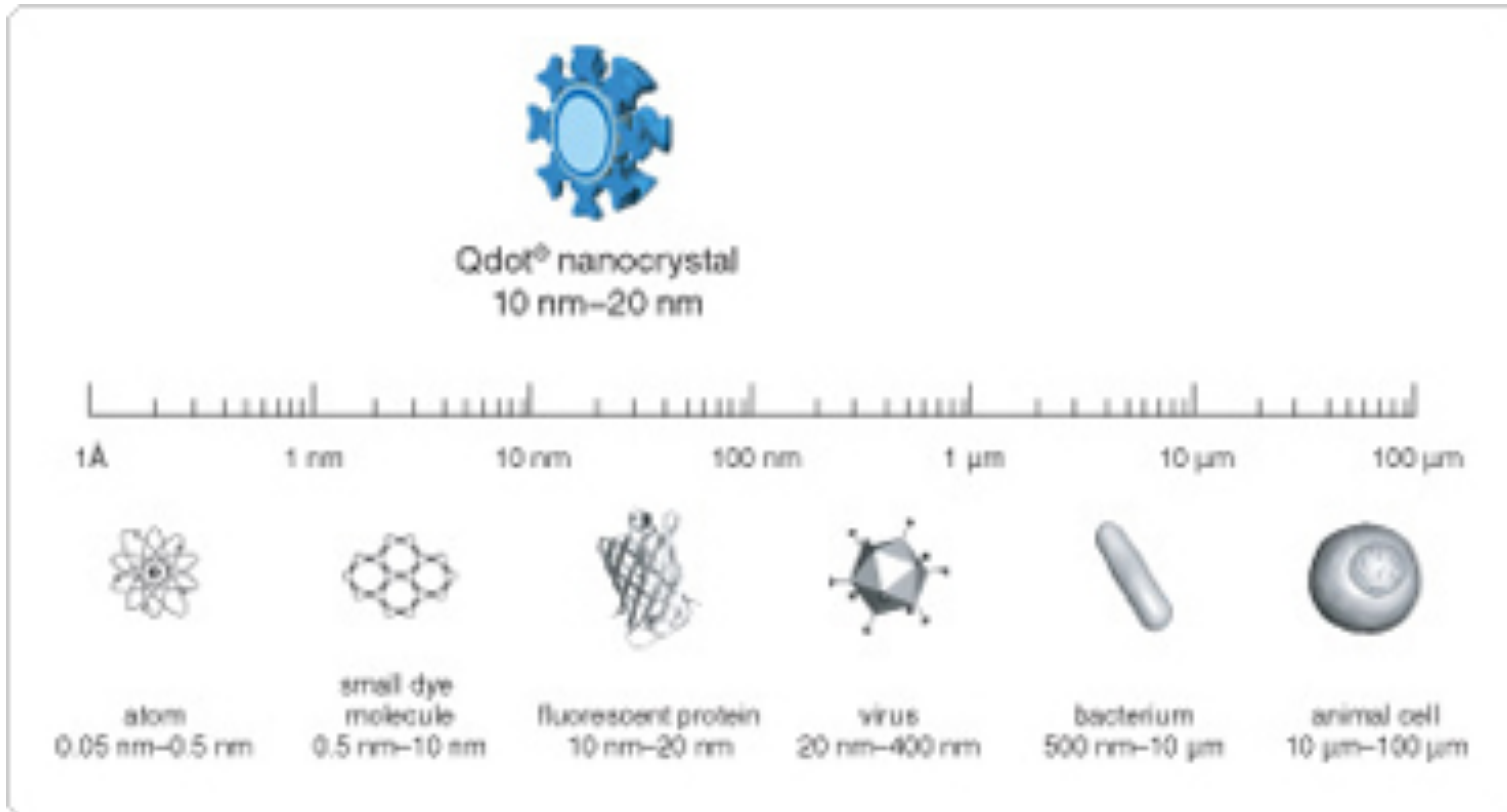


Core-Shell Semiconductor Nanocrystals (CdSe/ZnS)

Quantum-Confinement: Absorption and Emission is dependent upon size.

Due to the number of atoms- Many energy levels=continuous absorption/excitation.

Quantum-Dots



Labeling Biological Targets

Fixed Cells

Fluorescent In-Situ Hybridization (FISH)
uses target (epitope) specific antibody
(primary) to mark the target, and another
fluorescently labeled antibody (secondary)
which is specific against the first antibody

Live- Cells

Fluorescent-Proteins.

Genetically coded tags.

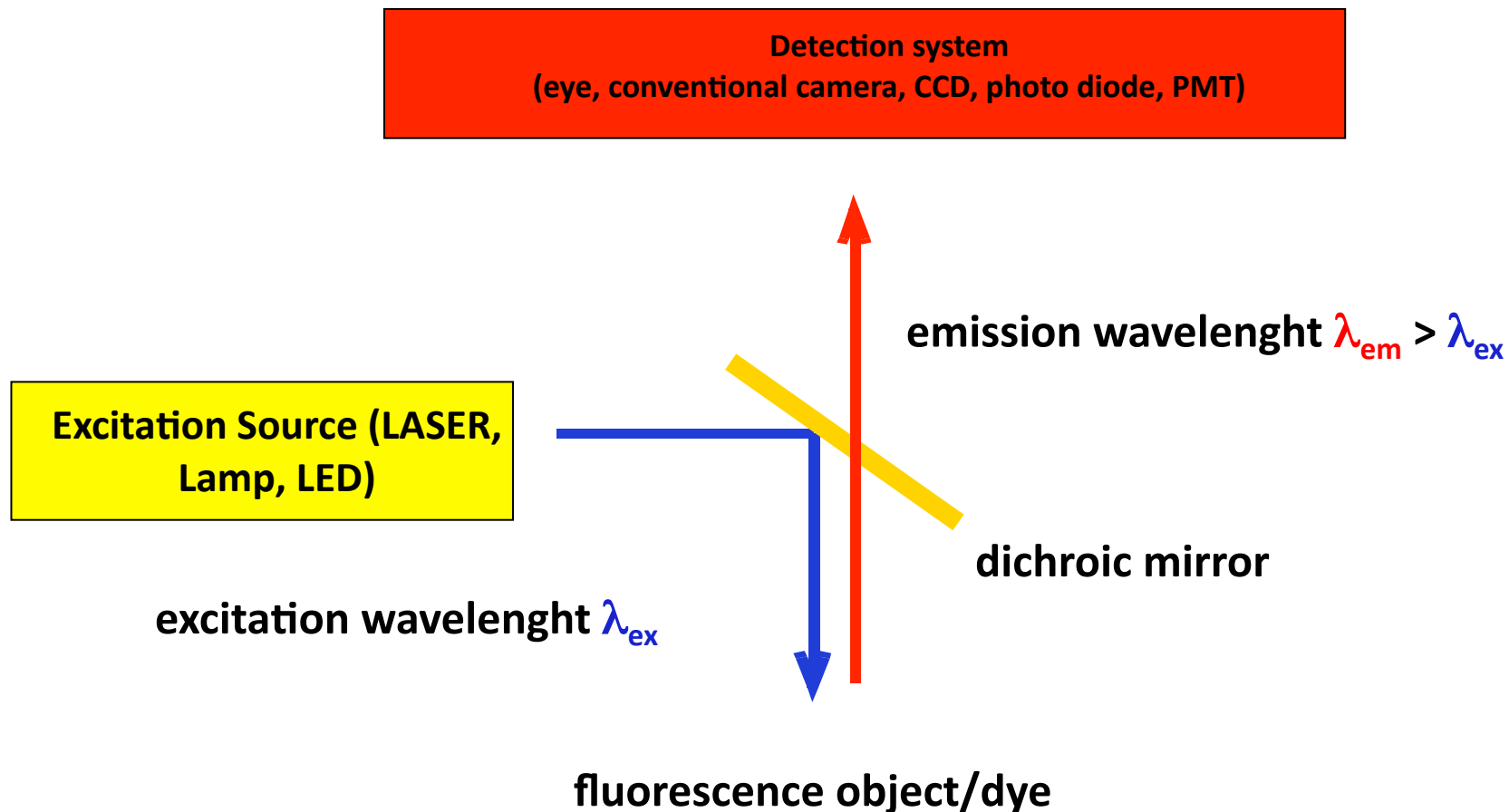
Cellular delivery of dyes: Membrane permeable
Chemically/electrically induced cell uptake.
Microinjection

Chemically specific Dyes.
Membrane Dyes, DNA, RNA intercalating dyes,
mitochondria dyes.

INSTRUMENTATION

- Excitation sources
- Filter sets
- Objectives
- Detection

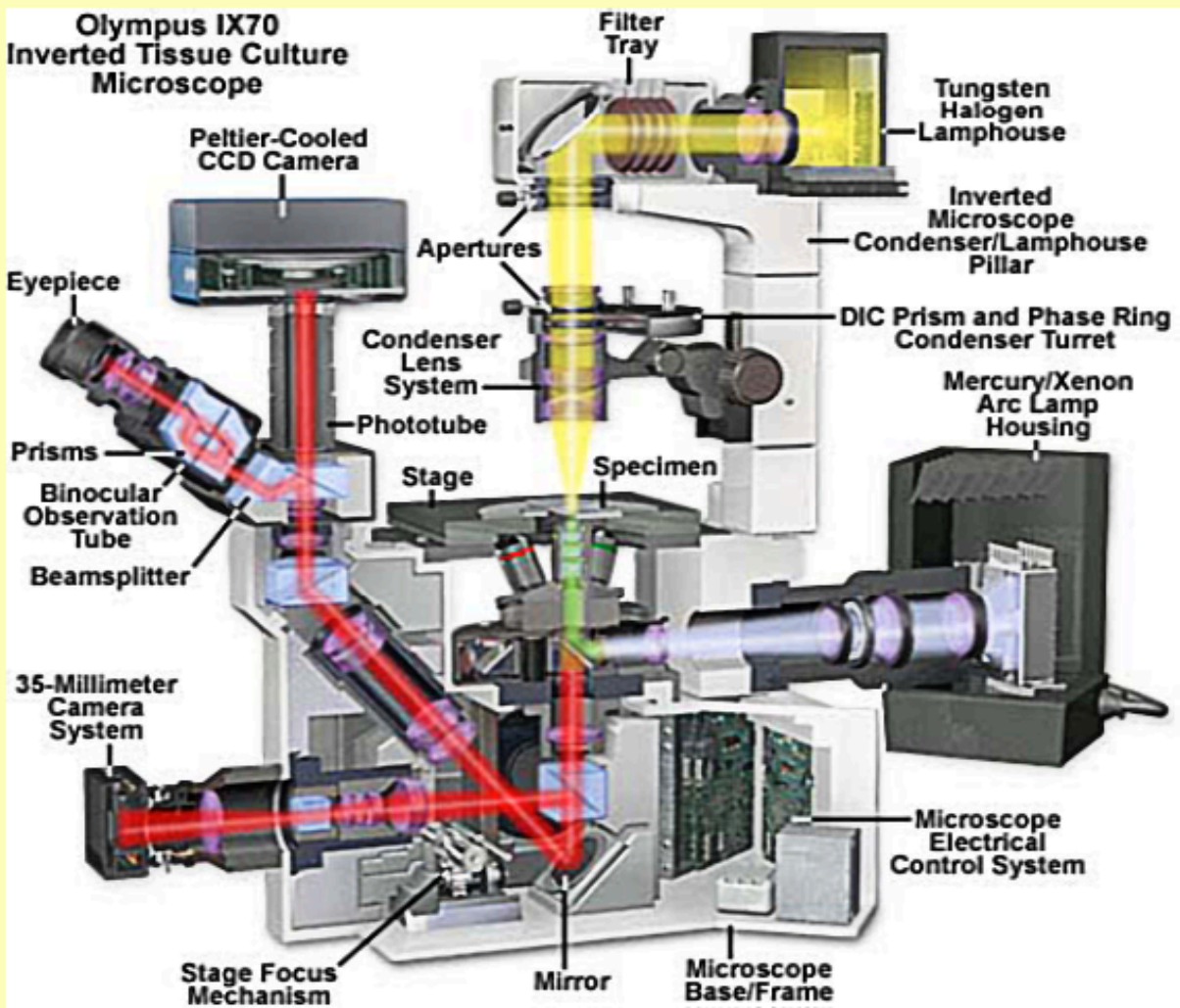
Fluorescence Microscopy



Excitation: Shine Light at one wavelength and get fluorescence at another (lower energy)

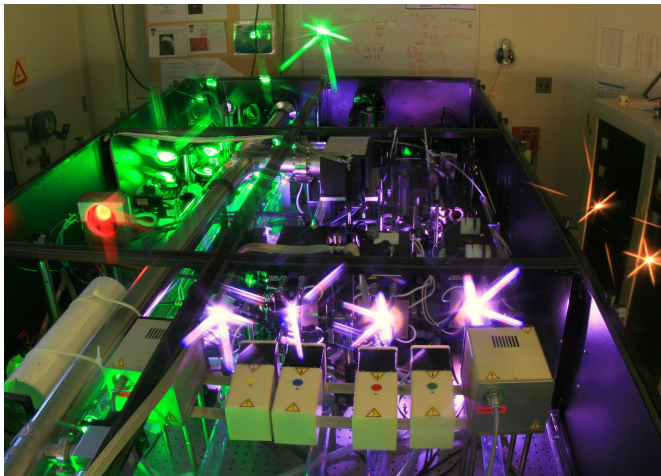
INVERTED MICROSCOPE

Olympus IX70
Inverted Tissue Culture
Microscope



Fluorescence Microscopy

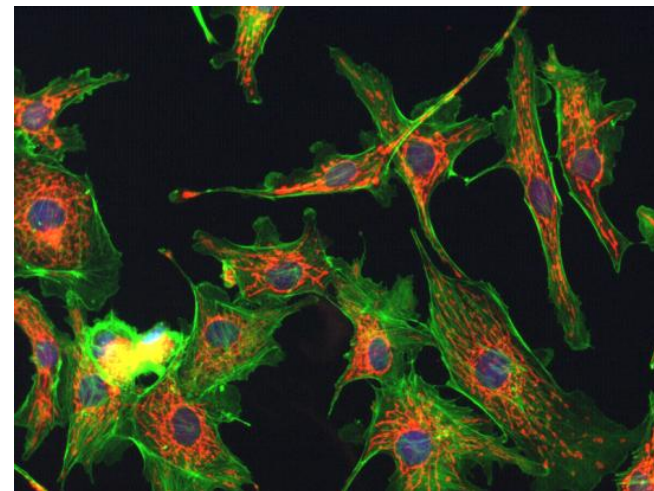
Excitation Source: Lasers, LEDs
(narrow wavelength), Lamps



Fluorophores (fluorescent molecules):

Quantum-Yield (probability of absorbed photon resulting in an emitted photon)

Absorption cross section (probability of absorbing a photon or how many photons we need to excite the molecule)



Excitation Sources

Arc lamps

Xenon

Mercury

Laser types



Argon	351	364	457	477	488	514
Blue diode		405	440			
Helium-Cadmium	354		442			
Krypton-Argon			488		569	647
Green Helium-Neon					543	
Yellow Helium-Neon						594
Orange Helium-Neon						612
Red Helium-Neon						633
Red diode						635 650
Ti:Sapphire					720-980	

FILTER CUBES

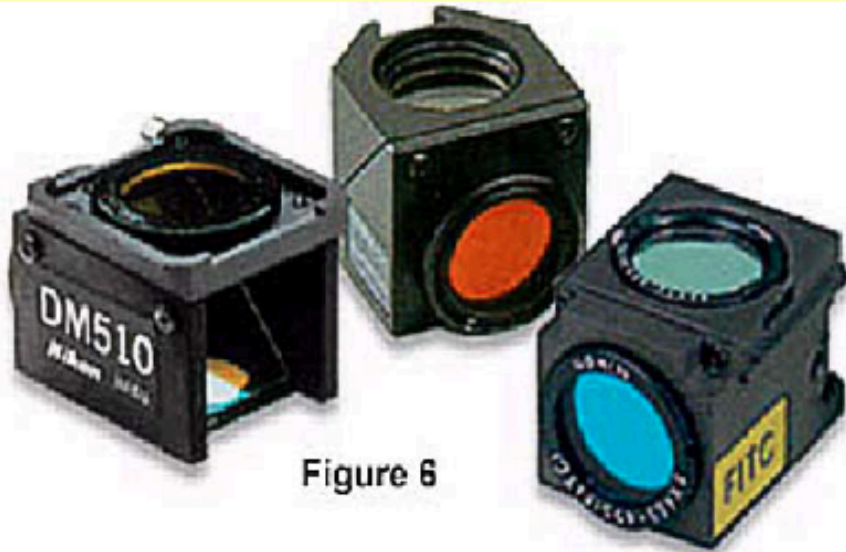


Figure 6

Fluorescence Interference Filter Block

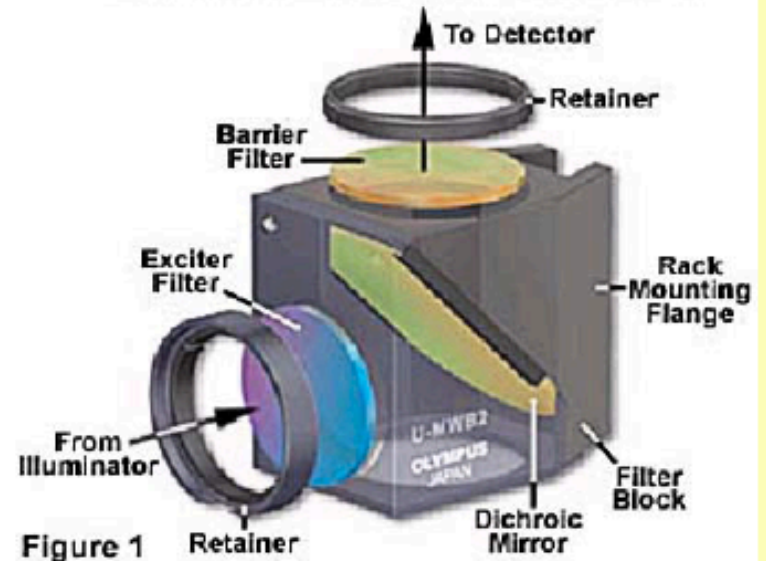
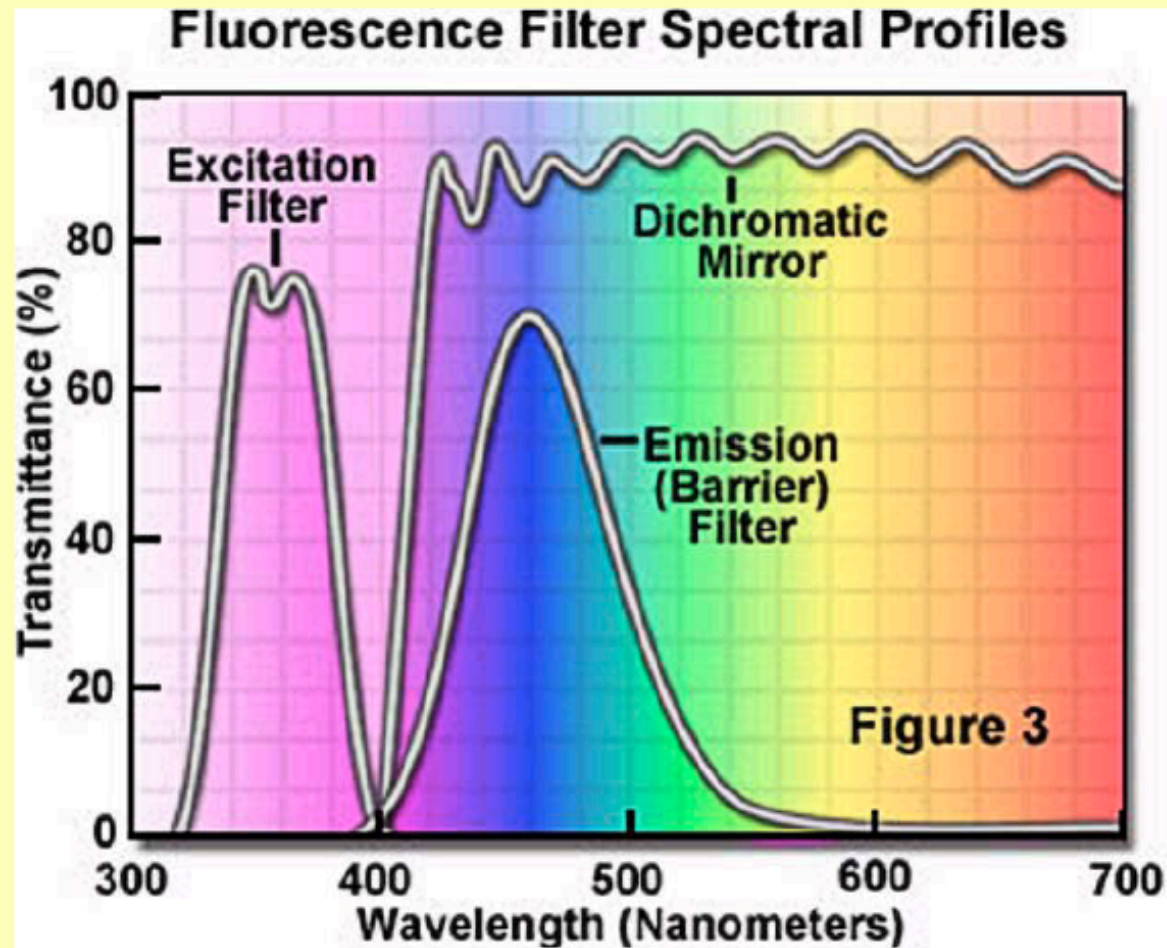


Figure 1

FILTER TRANSMISSION CURVES



Fluorescence Microscopy

Microscope Objective

(collecting the fluorescence from sample):

- High magnification (63x, 100x)
- High Numerical Aperture (N.A. > 1.4)

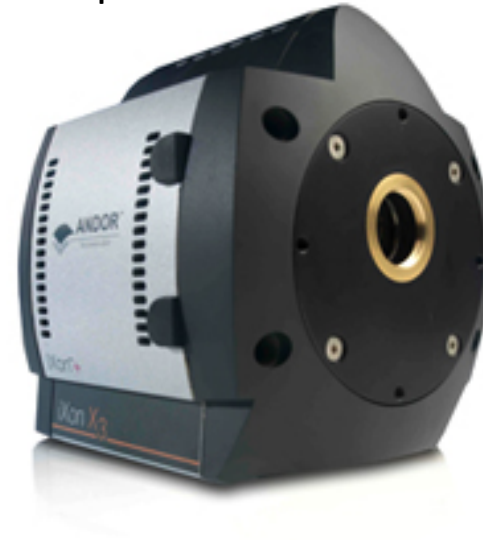


Detectors

EMCCD camera

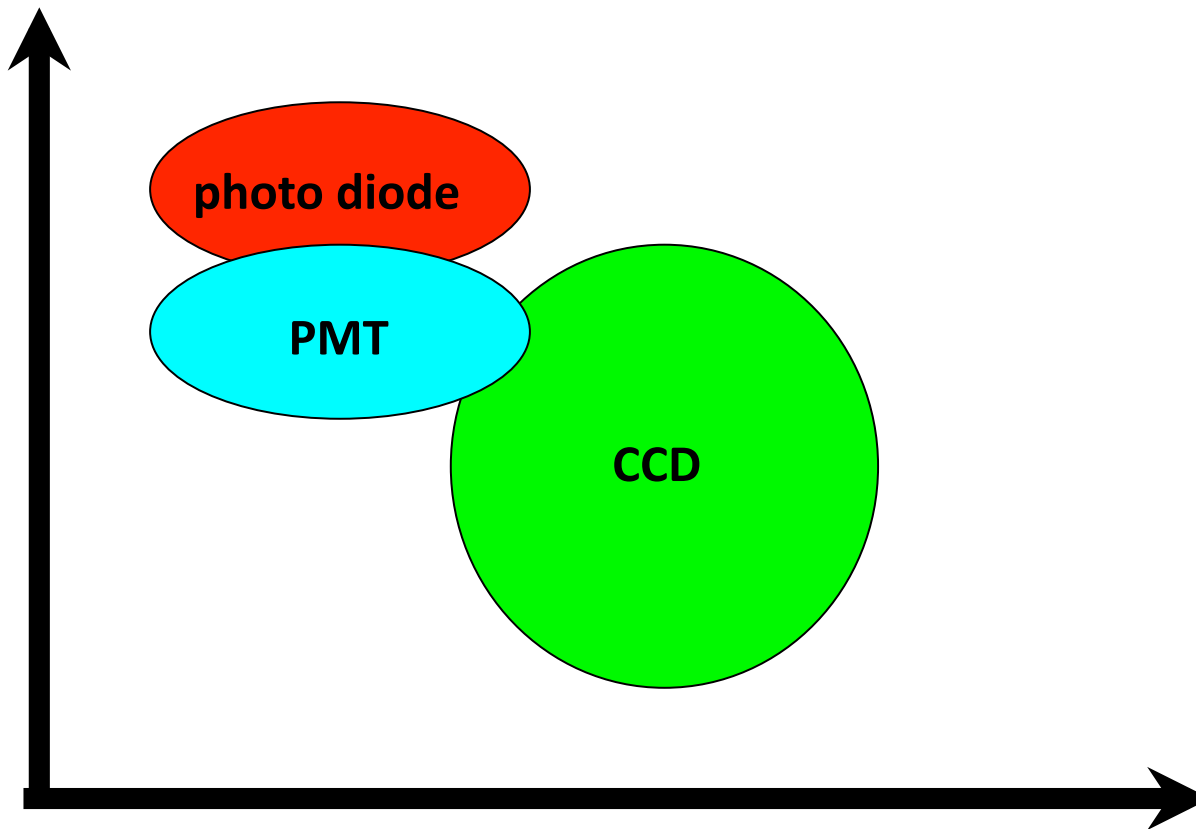
(electron multiplying Charge-Couple Device):

- High Quantum Efficiency (QE) – sensitivity to light
- Fast Acquisition rates



Fluorescence detector systems ...

Temporal
resolution



Spatial
resolution

OPTIMIZE DETECTION

Improve QE

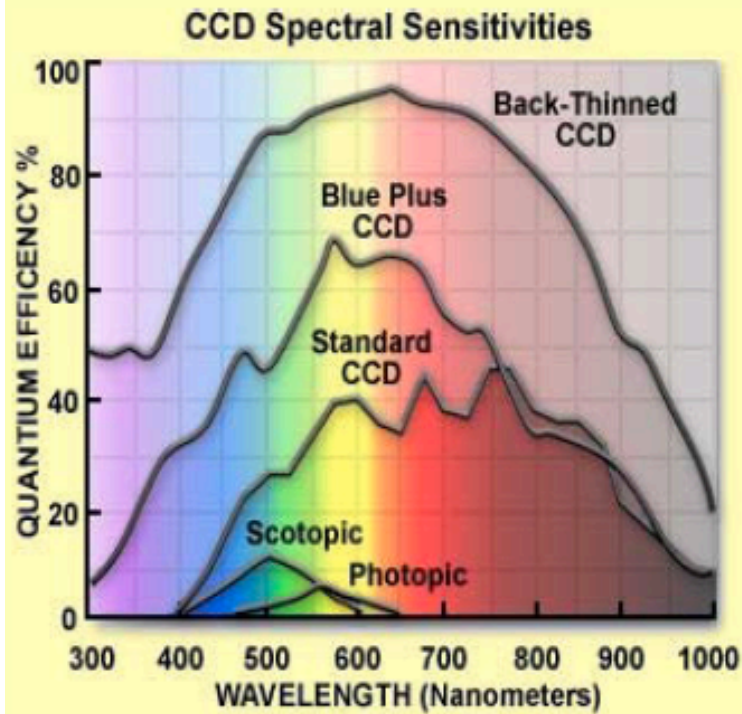
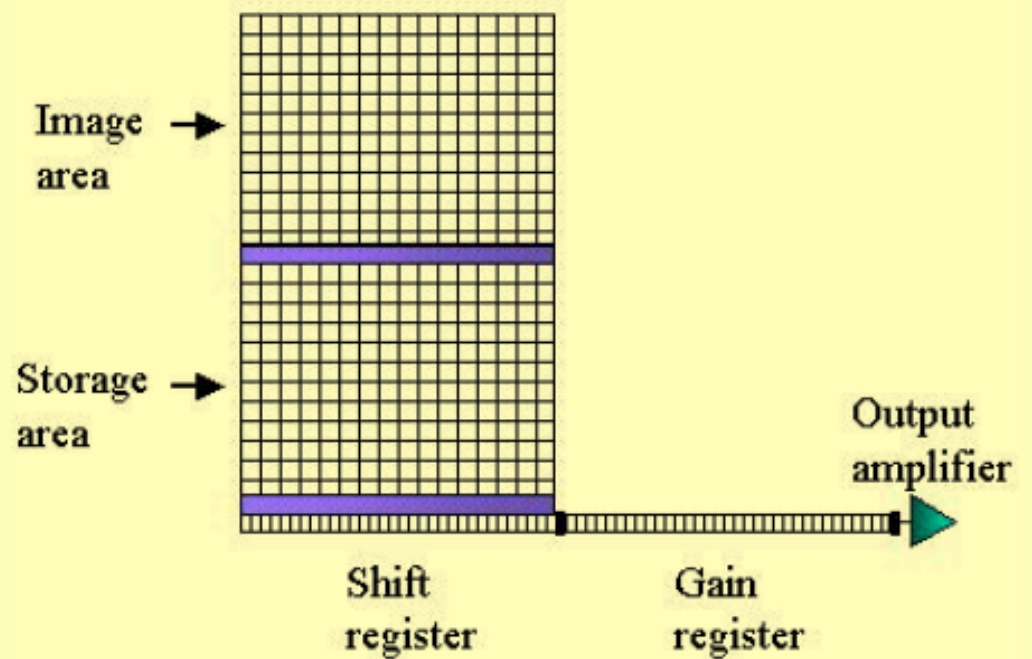
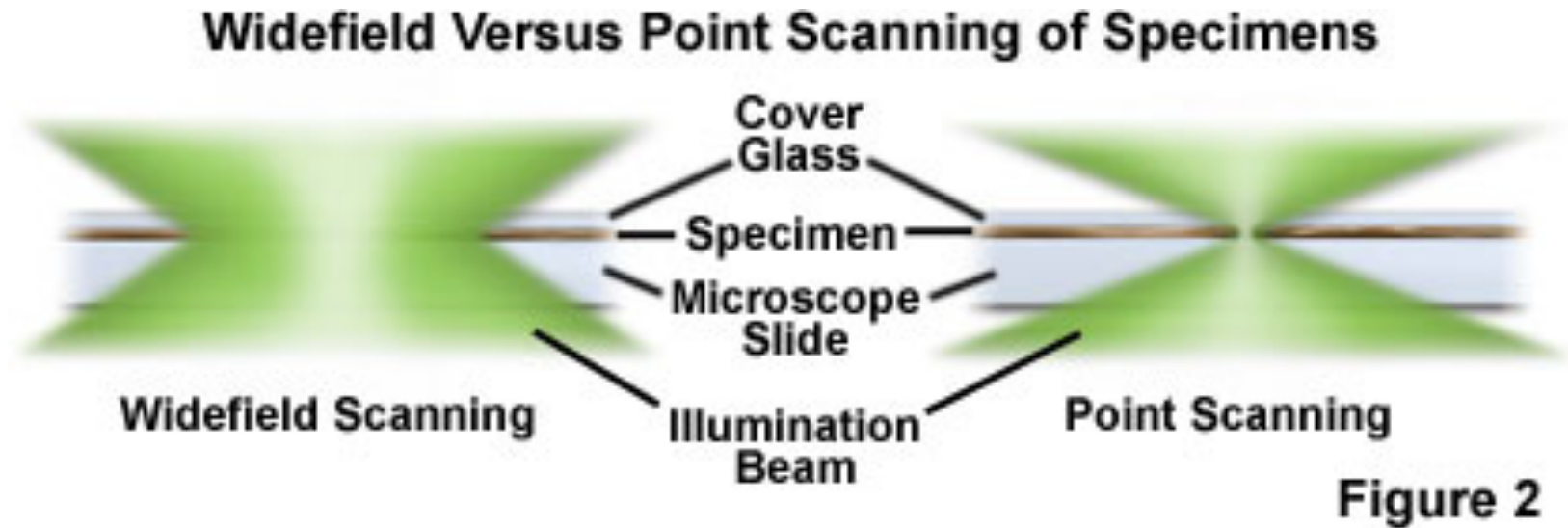


Figure 6

Reduce readout noise



Disadvantages of widefield microscopy



EFFECT OF NA ON RESOLUTION

